

Membrane Biochemistry

Lectures by

John F.Allen

School of Biological and Chemical Sciences, Queen Mary, University of London

jfallen.org/lectures



Queen Mary
University of London



Sunday, November 28, 2010

Bi-Visions

AT HARVARD UNIVERSITY

click to close



brought to you by
Bi-Visions

THE MITOCHONDRIA

WATCH THE EXCLUSIVE CLIP



click to play



Mr
2007

Further reading

- [Chemiosmotic coupling: The cost of living](#). By Peter Rich. (.pdf file, 80 kb)
- [Power for Life](#). Review of Nick Lane's book "Power Sex Suicide...." (.pdf file, 416 kb)
- [N,K-ATPase](#). Page of Mark Hilge at Protein Biophysics, Nijmegen
- [ATP Synthase](#). Group Pages of John Walker at the MRC Mitochondrial Biology Unit, Cambridge

Animations

- [The pump cycle of Na,K-ATPase](#). By Mark Hilge at Protein Biophysics, Nijmegen
- [Animation. From Light to ATP](#). By O. Fritzsche and W. Junge, University of Osnabruck. (.avi file, 17.7 mb)
- [Molecular animations of ATP synthase](#). From the research group of John Walker at the MRC Mitochondrial Biology Unit, Cambridge
- [Animation. Powering the Cell: Mitochondria](#). From BioVisions at Harvard University

Relevant Nobel prizes

- [1906 Nobel Prize in Physiology or Medicine to Camillo Golgi and Santiago Ramón y Cajal](#)
- [1974 Nobel Prize in Physiology or Medicine to Albert Claude, Christian de Duve and George E. Palade](#)
- [1978 Nobel Prize in Chemistry to Peter Mitchell](#)
- [1988 Nobel Prize in Chemistry to Johann Deisenhofer, Robert Huber and Hartmut Michel](#)
- [1997 Nobel Prize in Chemistry to Paul D. Boyer, John E. Walker and Jens C. Skou](#)
- [1999 Nobel Prize in Physiology or Medicine to Günter Blobel](#)



Lectures in Membrane Biochemistry

- The endomembrane system - endocytosis and exocytosis (Acrobat, .pdf file)
- The endomembrane system - vesicular transport and protein trafficking (Acrobat, .pdf file)
- Transport across membranes 1 - Proteins (Acrobat, .pdf file)
- Transport across membranes 2 - Small molecules and ions (Acrobat, .pdf file)
- Mitochondria and chloroplasts - analysis of compartments (Acrobat, .pdf file)
- Bioenergetics (Acrobat, .pdf file)
- The respiratory chain and oxidative phosphorylation (Acrobat, .pdf file)
- Oxidative phosphorylation, respiratory control and the chemiosmotic hypothesis (Acrobat, .pdf file)
- ATP Synthase - Coupling ATPase (Acrobat, .pdf file)

Course web pages

Membrane Biochemistry web pages

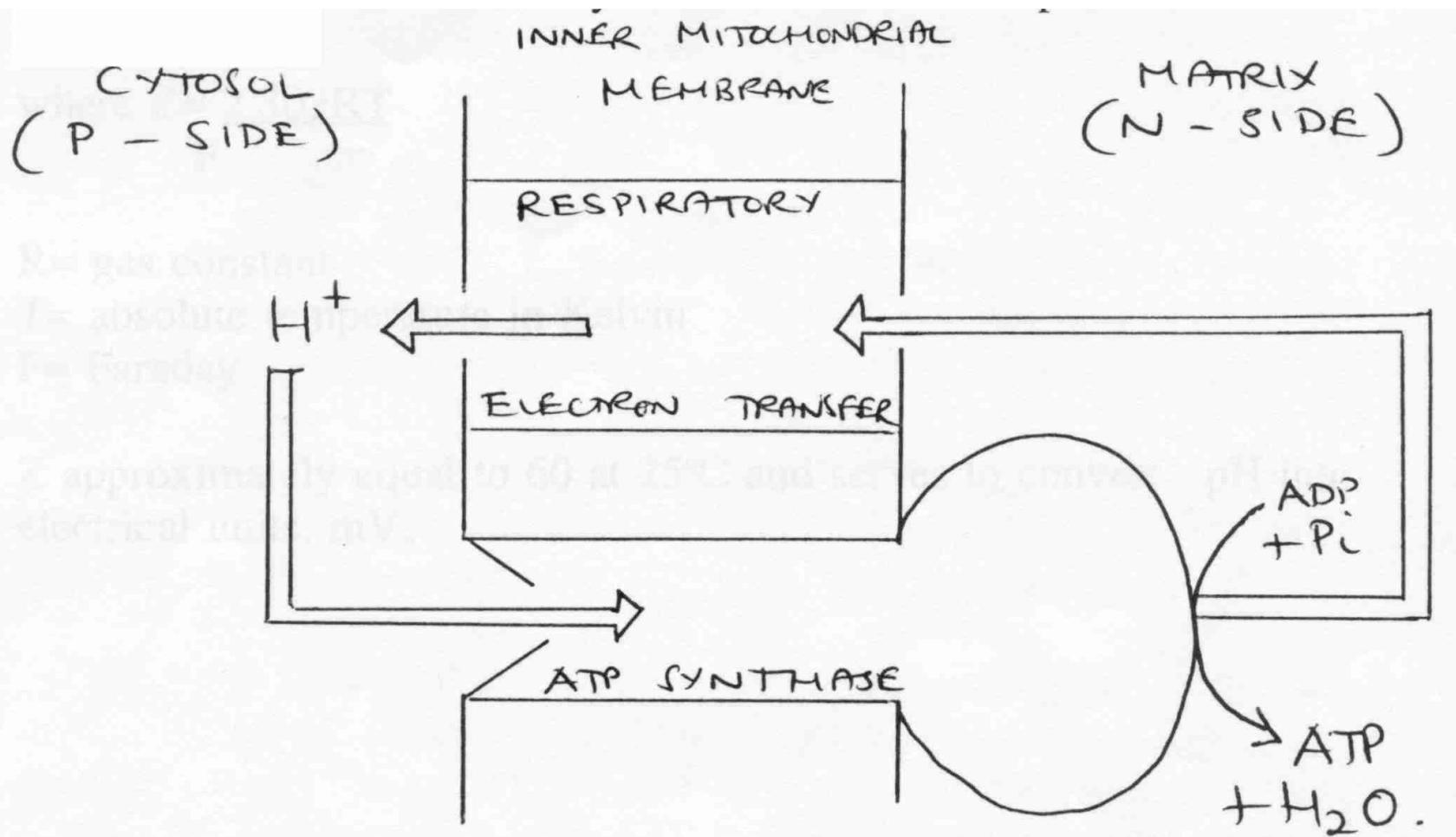
General reference

Cell and Molecular Biology: Concepts and Experiments
Gerald Karp. Fifth Edition 2008. John Wiley & Sons Inc.

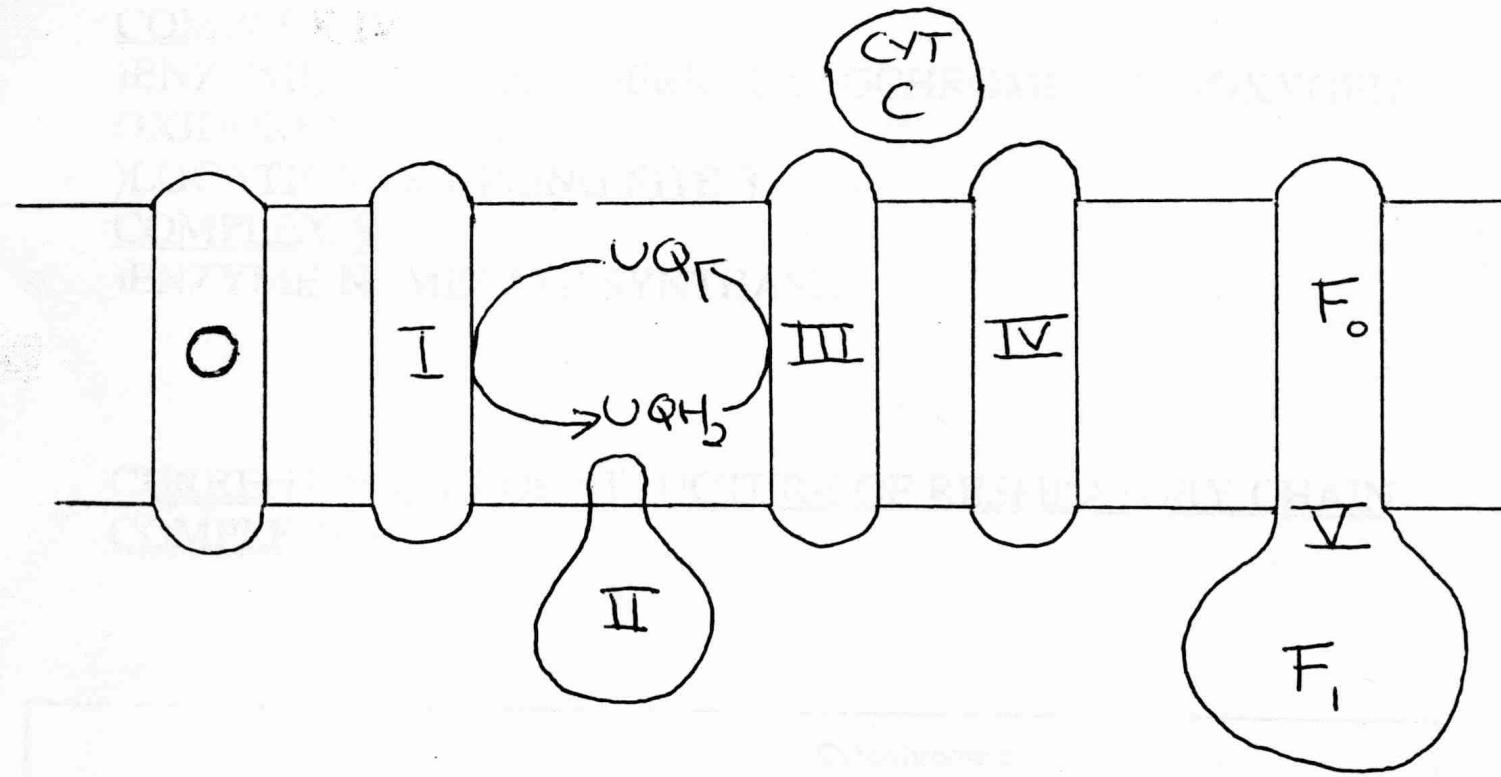
ATP Synthase – Coupling ATPase

Transmembrane Proton gradient

Energy transduction occurs by means of a proton circuit through the insulating, coupling membrane and between the two bulk aqueous phases (the matrix and the intermembrane space/cytosol in mitochondria).



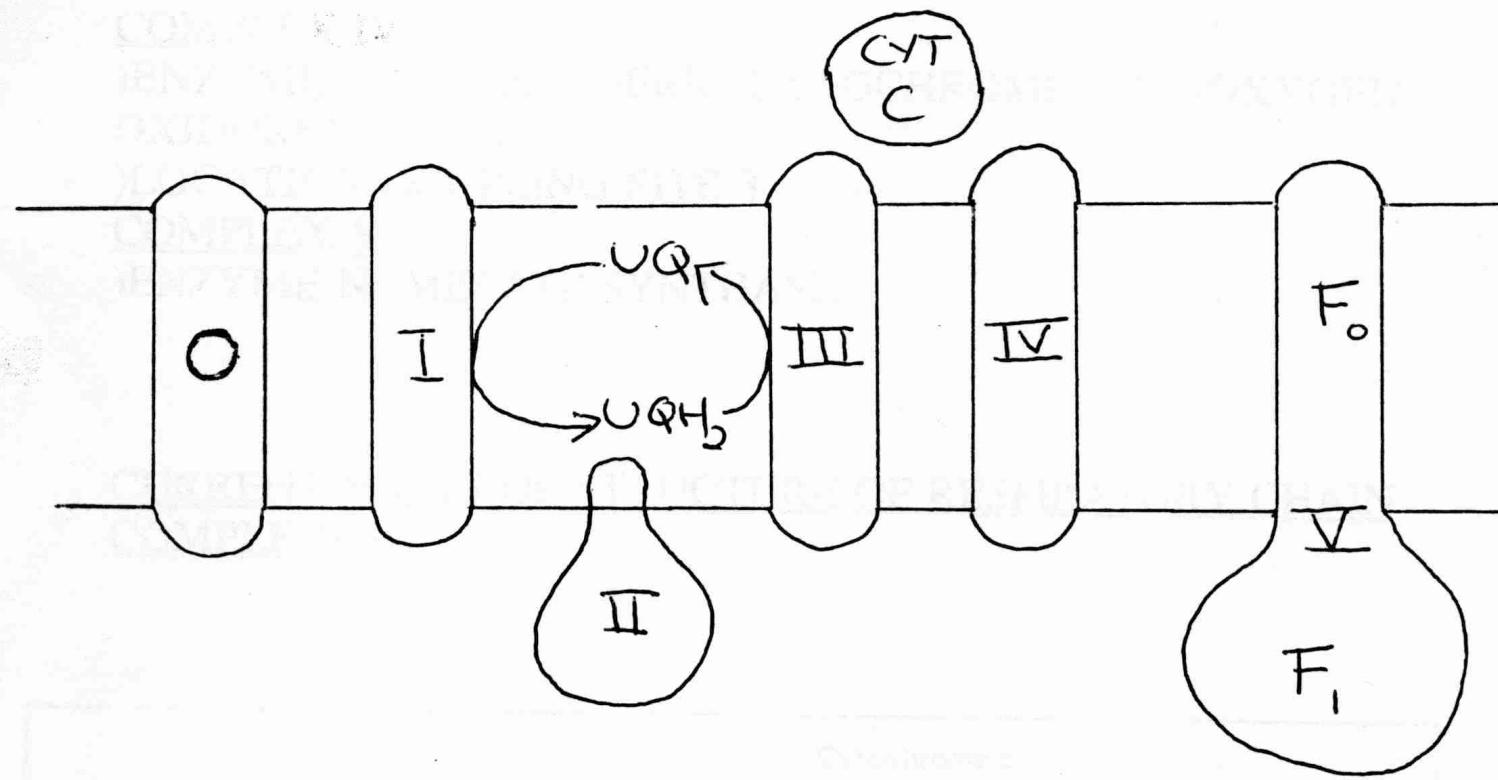
INTERMEMBRANE SPACE (OUTSIDE)



MITOCHONDRIAL MATRIX (INSIDE)

COMPLEX V

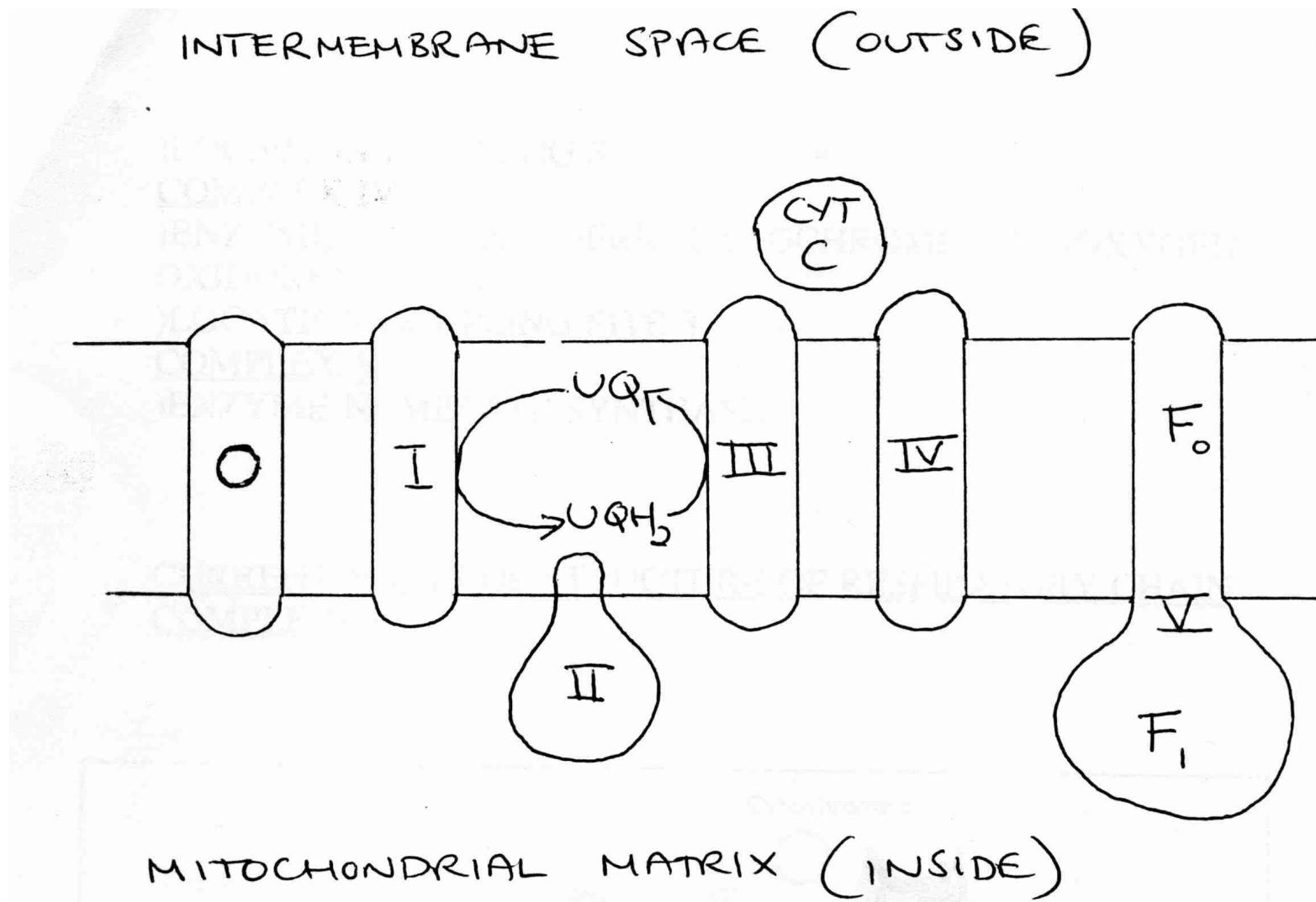
INTERMEMBRANE SPACE (OUTSIDE)



MITOCHONDRIAL MATRIX (INSIDE)

COMPLEX V

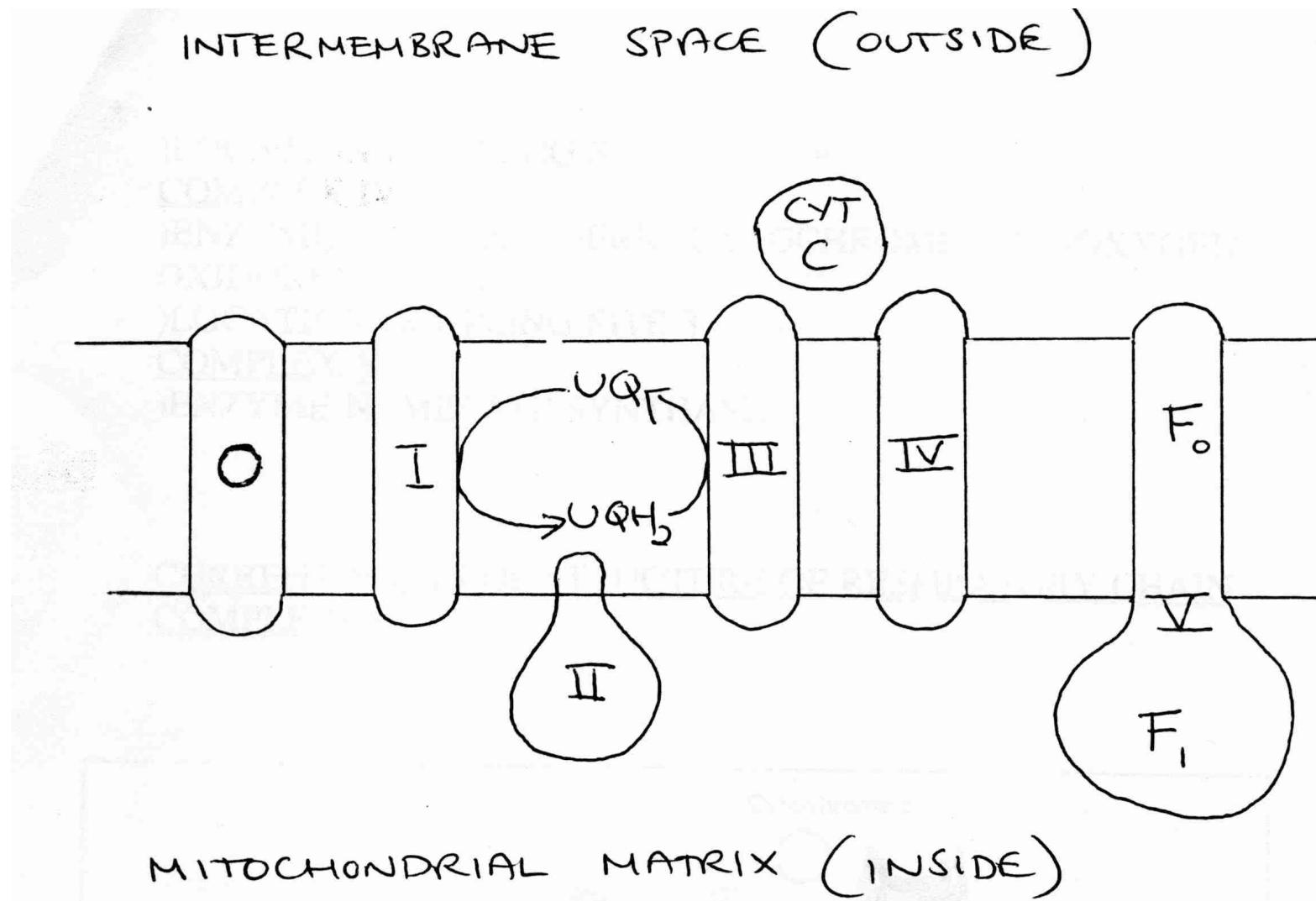
ENZYME NAME: COUPLING ATPase; ATP SYNTHASE



COMPLEX V

ENZYME NAME: COUPLING ATPase; ATP SYNTHASE

F-ATPase

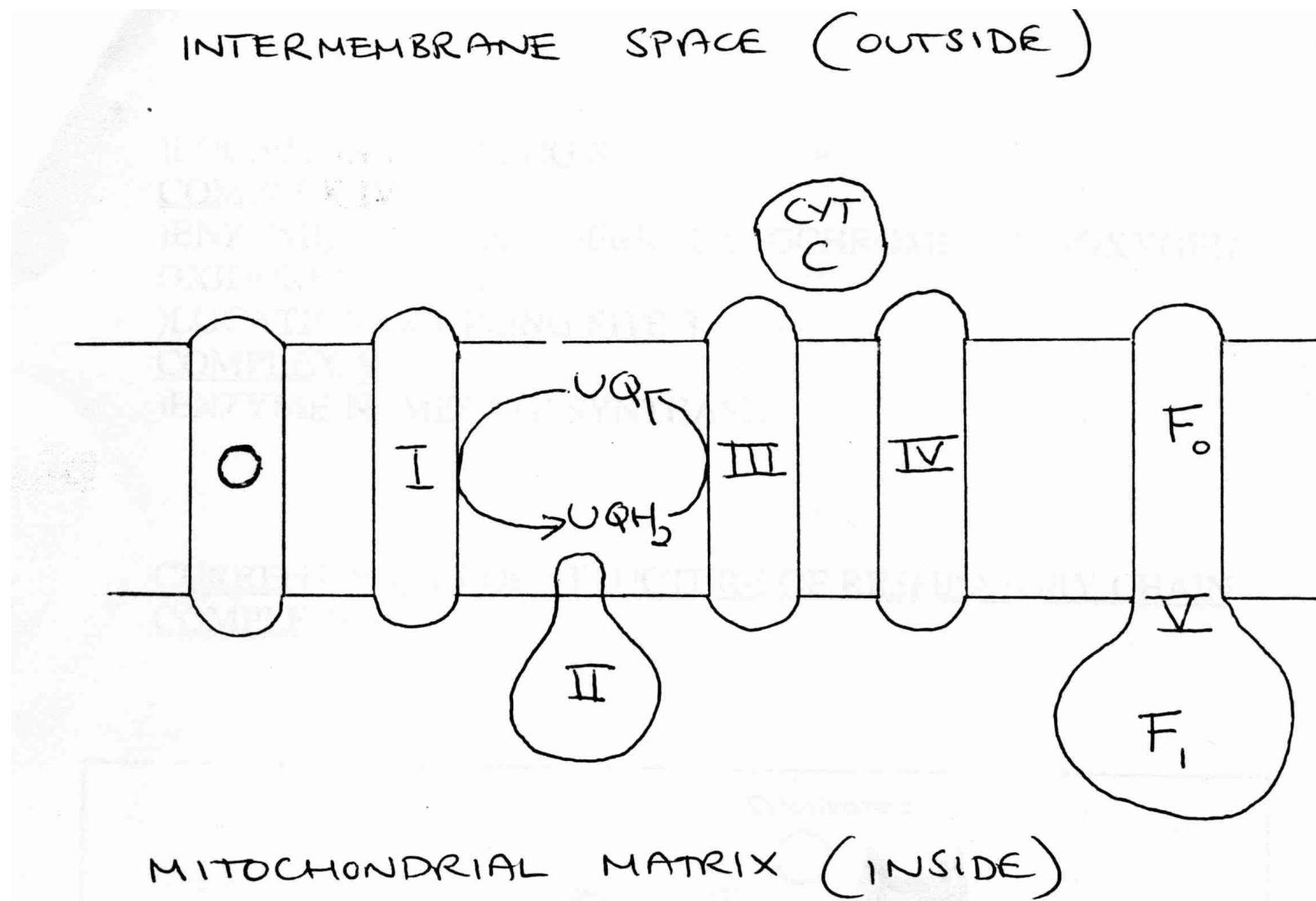


COMPLEX V

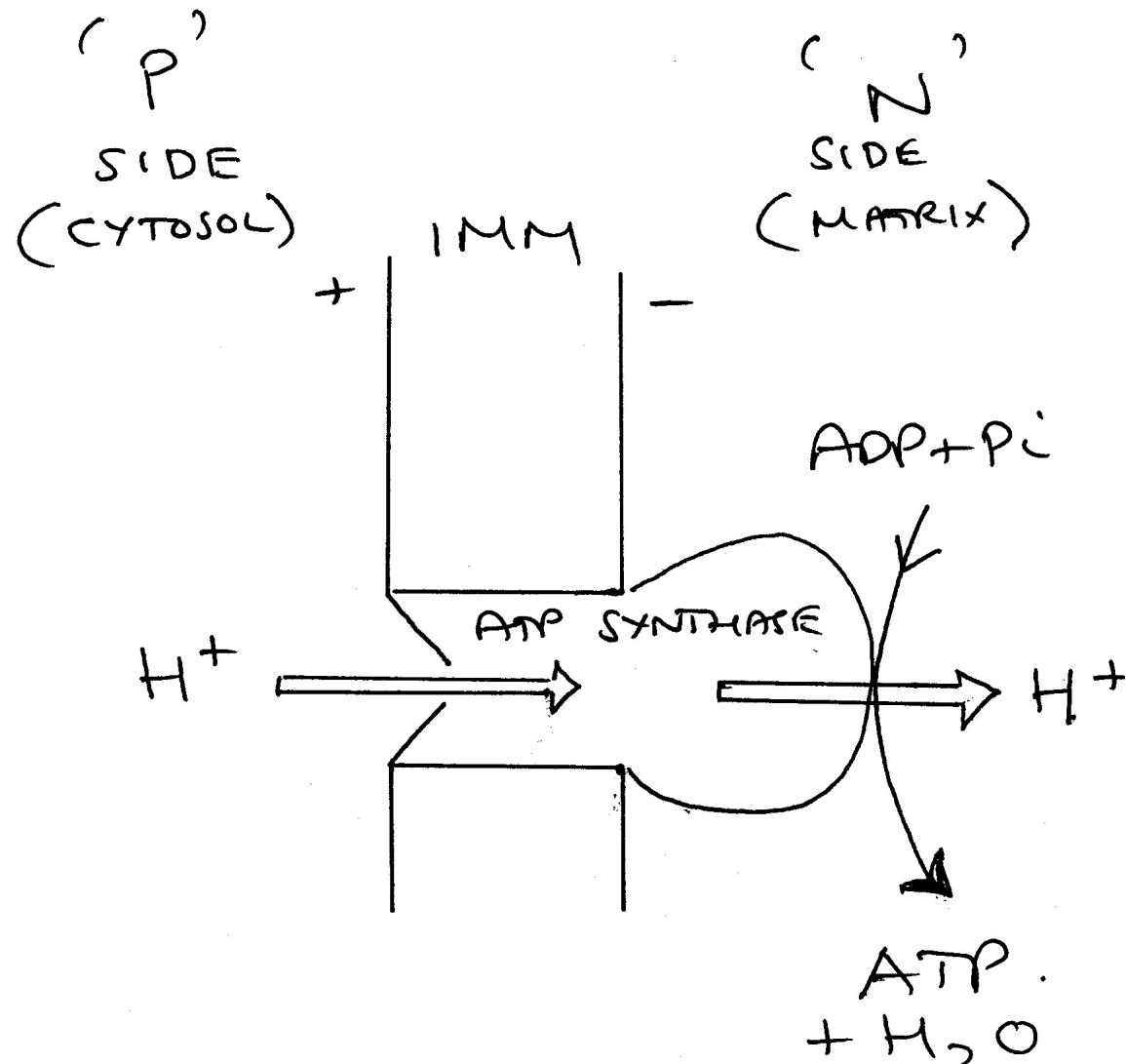
ENZYME NAME: COUPLING ATPase; ATP SYNTHASE

F-ATPase

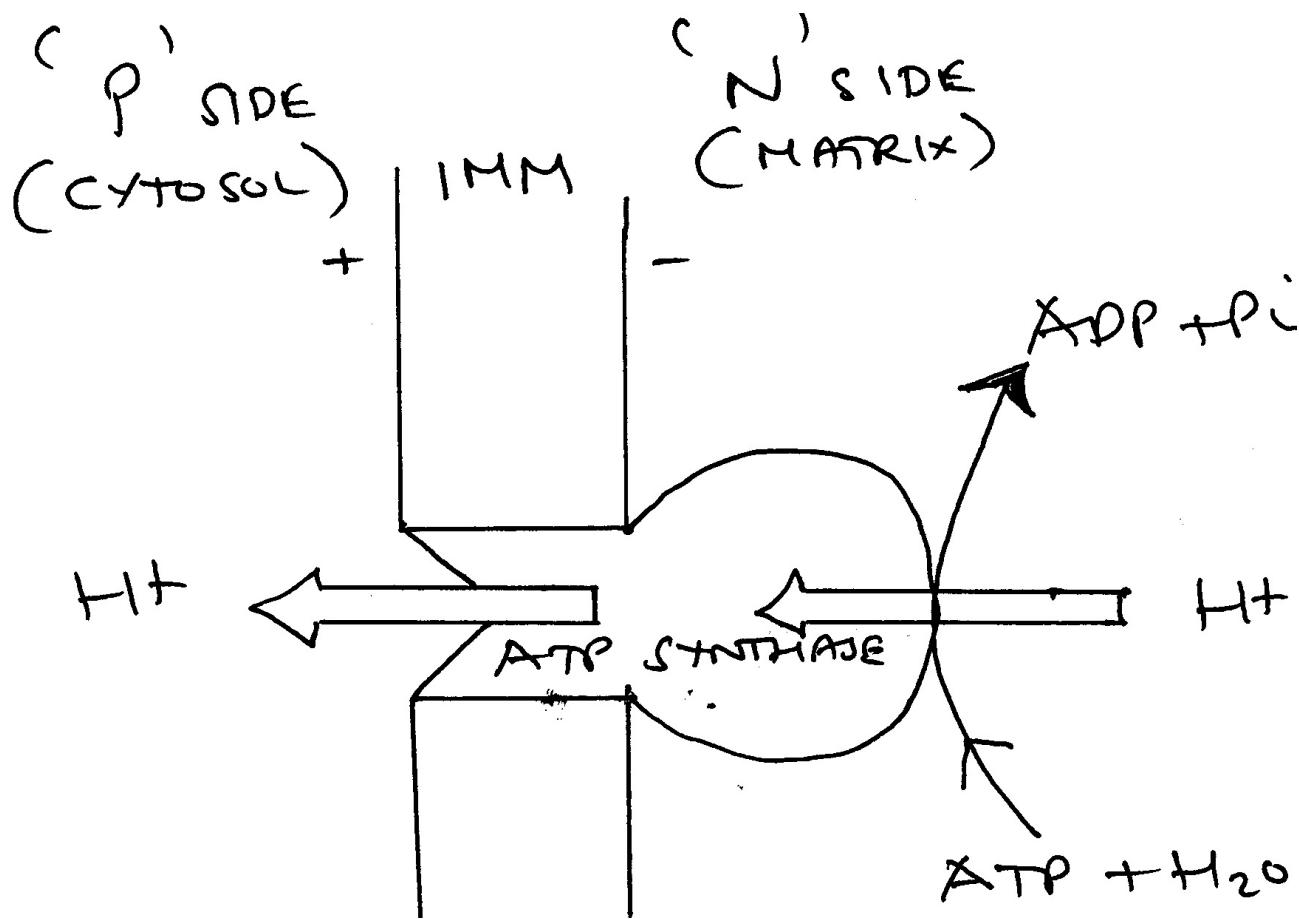
F₁-F₀ ATPase



ATP synthase can transport protons across the membrane in one direction down the concentration (ΔpH) and charge ($\Delta \psi$) gradient, using the energy for ATP synthesis.



Alternatively the ATP synthase can use the energy from ATP hydrolysis to pump protons in the opposite direction (active transport against the concentration and charge gradient).



Evidence for ΔpH -driven ATP synthesis

1. The 'acid-bath experiment'. ΔpH driving ATP synthesis was demonstrated in 1966 by Jagendorf and Uribe, using thylakoid membranes of chloroplasts.

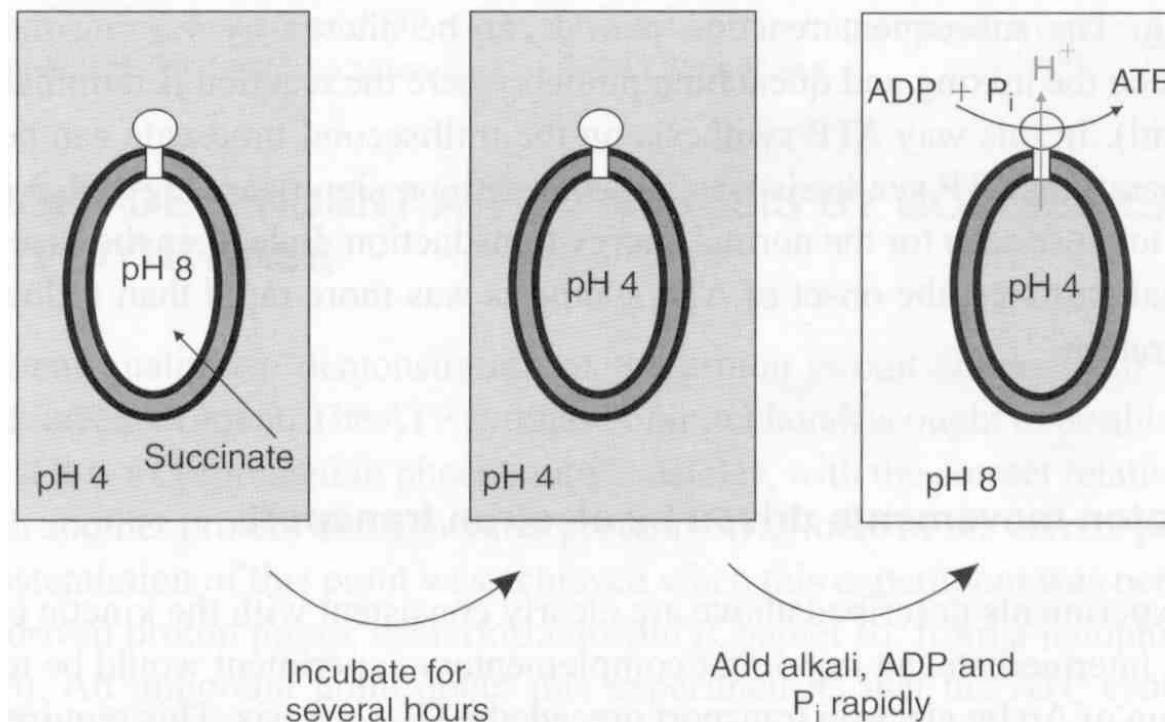
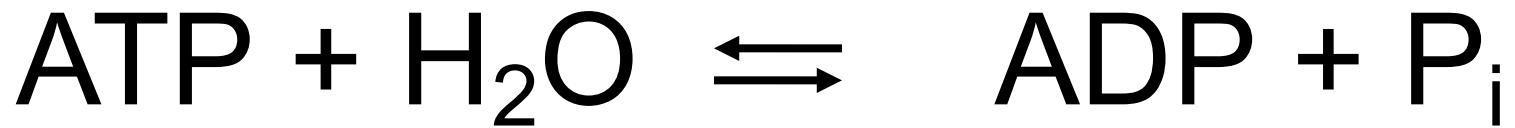


Figure 4.15 The 'acid bath' experiment: a ΔpH can generate ATP.

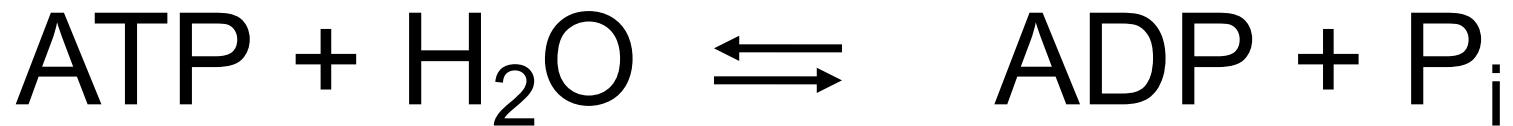
Thylakoid membranes were incubated in the dark at pH 4 in the presence of electron transport inhibitors in a medium containing succinate, which slowly permeated into the thylakoid space liberating protons and lowering the internal pH to about 4. The external pH was then suddenly raised to 8, creating a ΔpH of 4 units across the membrane. Traditionally H^+ efflux through the ATP synthase has been regarded as charge compensated by Cl^- efflux and/or Mg^{2+} influx. A $\Delta\psi$ may also be induced, see text. ADP and P_i were simultaneously added and proton efflux through the ATP synthase led to the synthesis of about 100 mol of ATP per mol of synthase. Protonophores such as FCCP inhibited the ATP production.

Coupling ATPase

Coupling ATPase

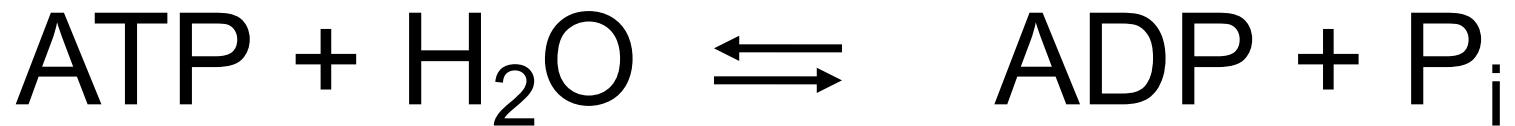


Coupling ATPase



Also known as “ATP Synthase”

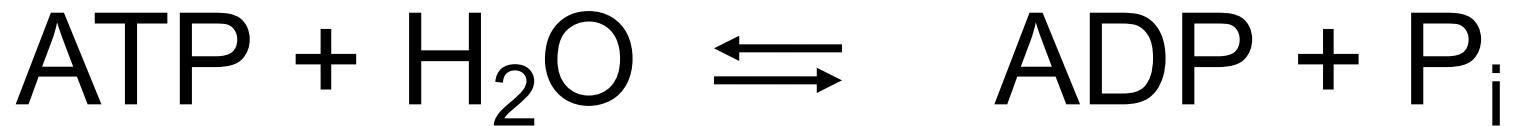
Coupling ATPase



Also known as “ATP Synthase”

One of the class of F-ATPases (c.f. V-ATPases; P-ATPases)

Coupling ATPase

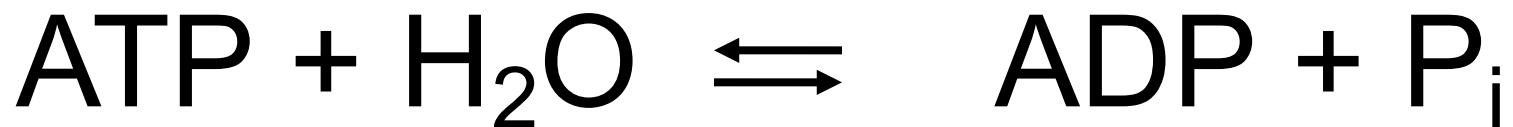


Also known as “ATP Synthase”

One of the class of F-ATPases (c.f. V-ATPases; P-ATPases)

- F₁

Coupling ATPase

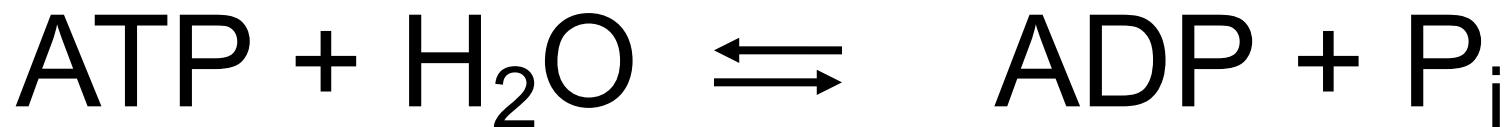


Also known as “ATP Synthase”

One of the class of F-ATPases (c.f. V-ATPases; P-ATPases)

- F_1
- F_o

Coupling ATPase



Also known as “ATP Synthase”

Also known previously as respiratory “complex V”

One of the class of F-ATPases (c.f. V-ATPases; P-ATPases)

- F_1
- F_o

F₁

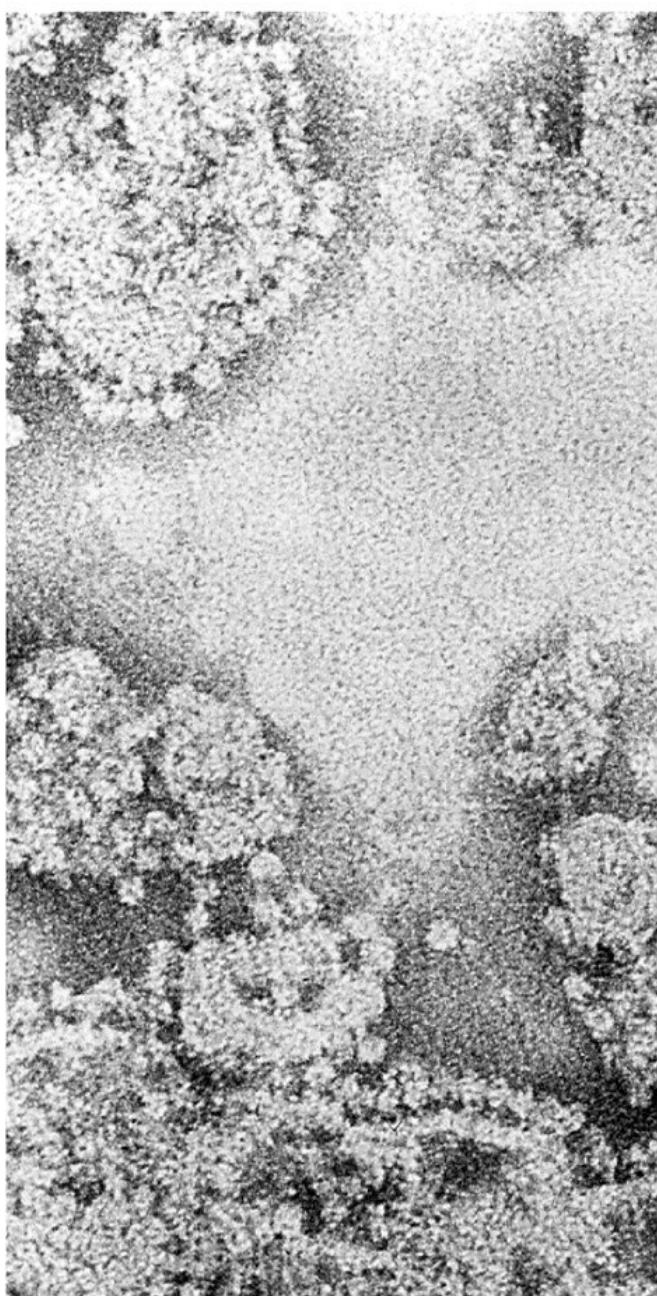
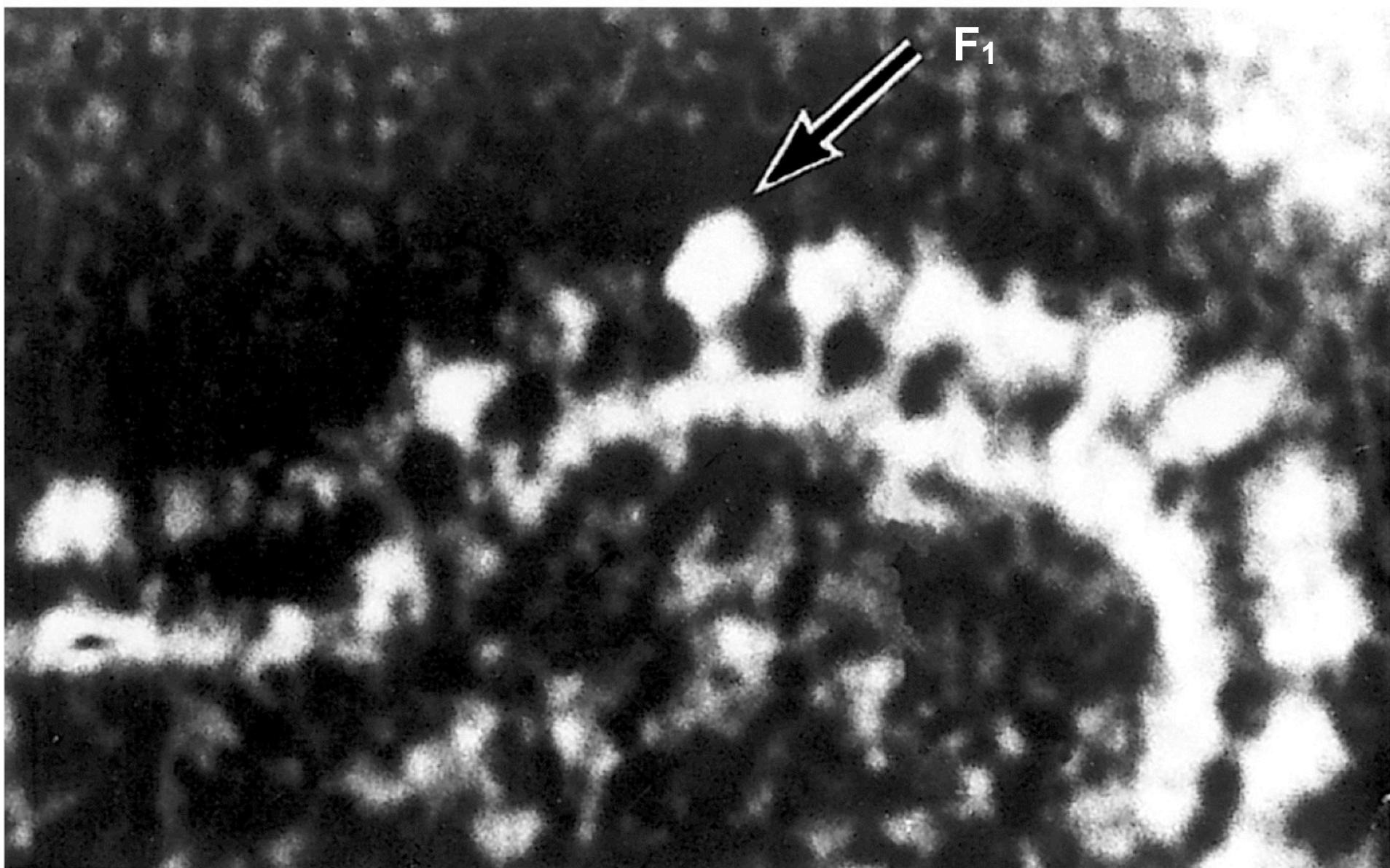


Figure 5-25a Cell and Molecular Biology, 4/e (© 2005 John Wiley & Sons)



10 nm

Figure 5-21 Cell and Molecular Biology, 4/e (© 2005 John Wiley & Sons)

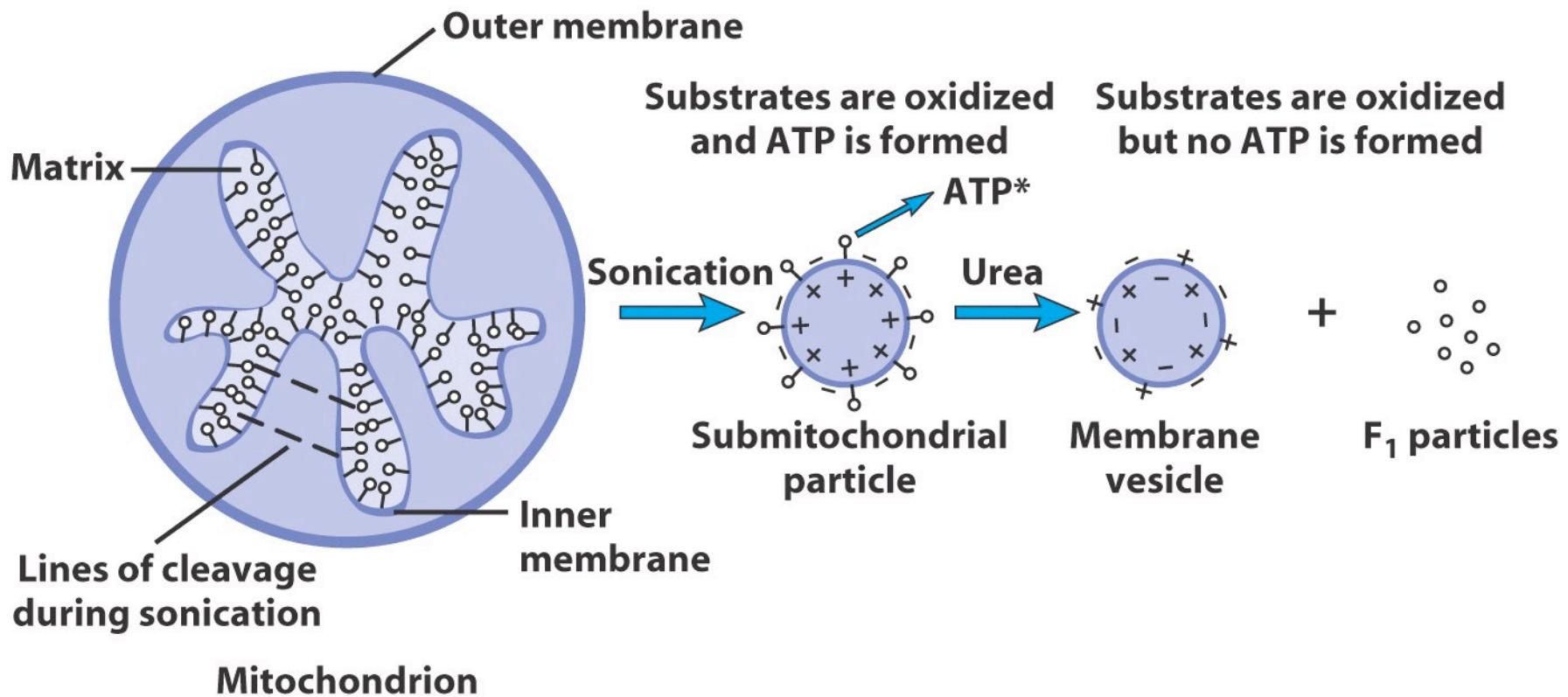


Figure 5-25b Cell and Molecular Biology, 4/e (© 2005 John Wiley & Sons)

Evidence for ΔpH -driven ATP synthesis

2. Experiment of Racker and co-workers, demonstrating light-driven ATP synthesis in lipid vesicles containing bacteriorhodopsin (a light-drive proton pump) and ATP synthase from mitochondria from beef heart

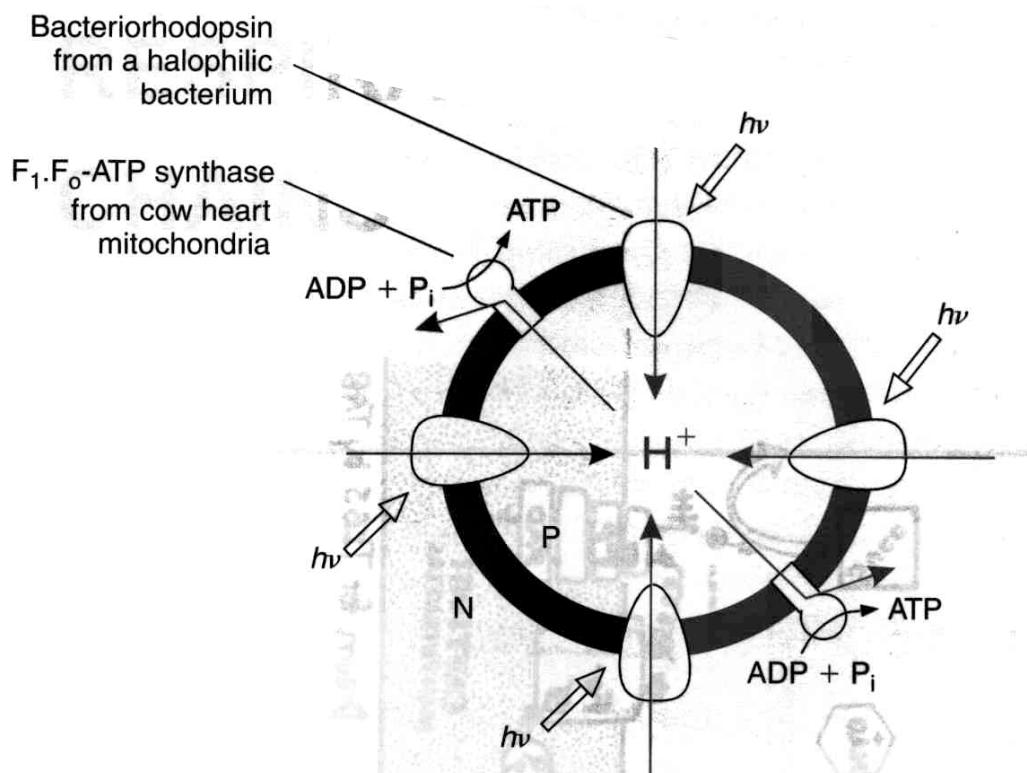


Figure 4.16 A proton circuit between a light-driven proton pump (bacteriorhodopsin) and ATP synthase from mitochondria.

The establishment of the proton circuit depends on the majority of the bacteriorhodopsin molecules adopting (for poorly understood reasons) the orientation in which they pump protons inward. Similarly, the ATP synthase had to incorporate predominantly with the topology shown. Opposite orientations of both bacteriorhodopsin and ATP synthase would in principle also have permitted an H^+ circuit, in the opposite direction, to be established. In practice, this would have meant that added ADP and P_i (both membrane impermeant) would not have been able to reach the active site of the ATP synthase.

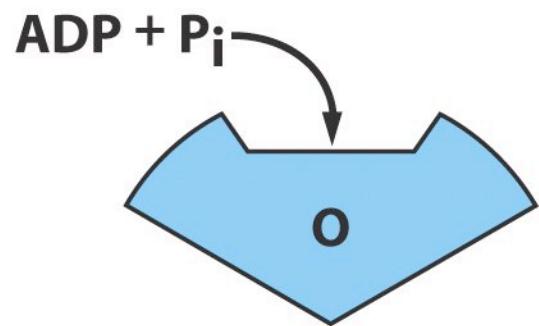


Figure 5-27a part 1 Cell and Molecular Biology, 4/e (© 2005 John Wiley & Sons)

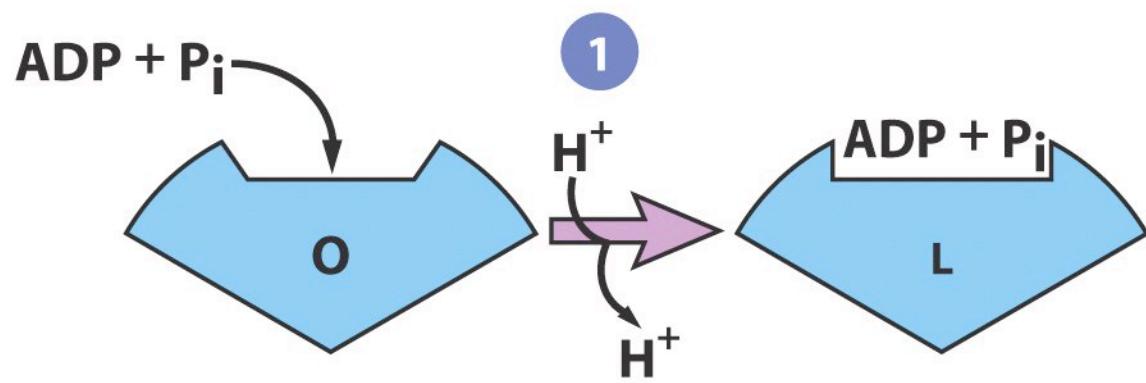


Figure 5-27a part 1 Cell and Molecular Biology, 4/e (© 2005 John Wiley & Sons)

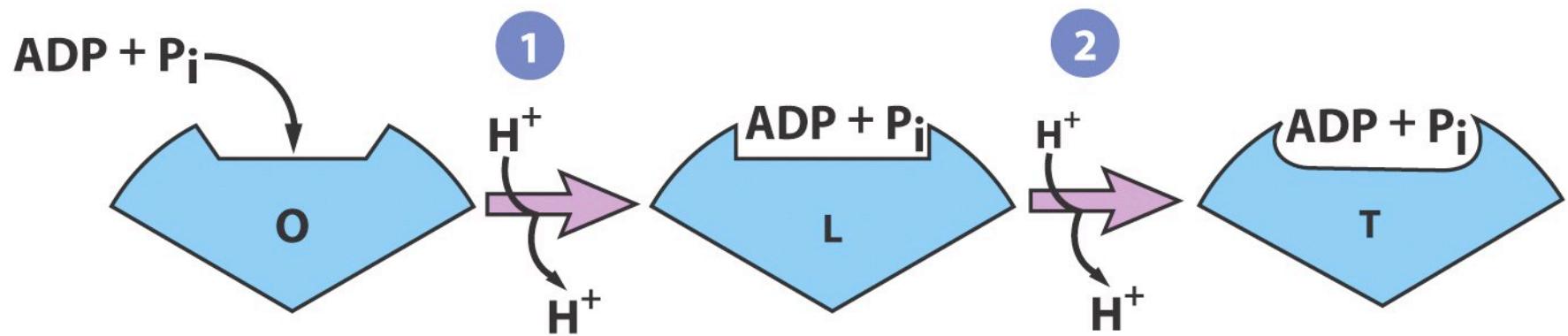


Figure 5-27a part 1 Cell and Molecular Biology, 4/e (© 2005 John Wiley & Sons)

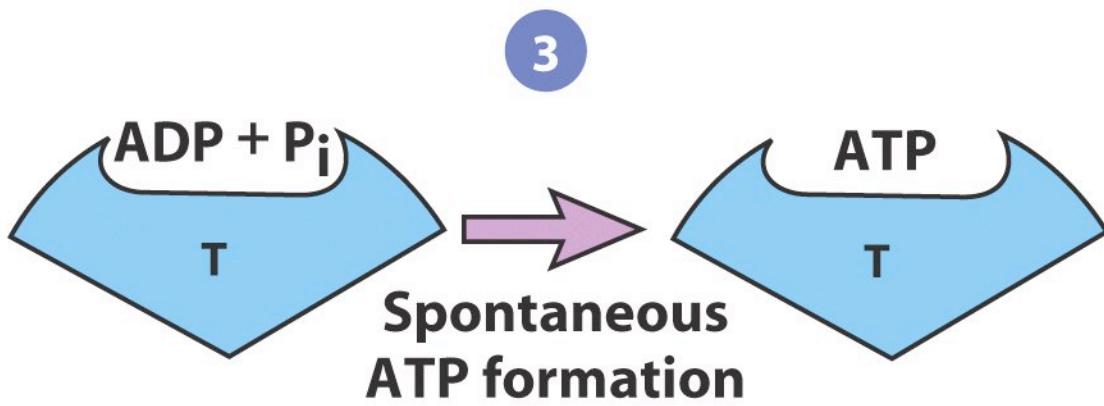


Figure 5-27a part 2 Cell and Molecular Biology, 4/e (© 2005 John Wiley & Sons)

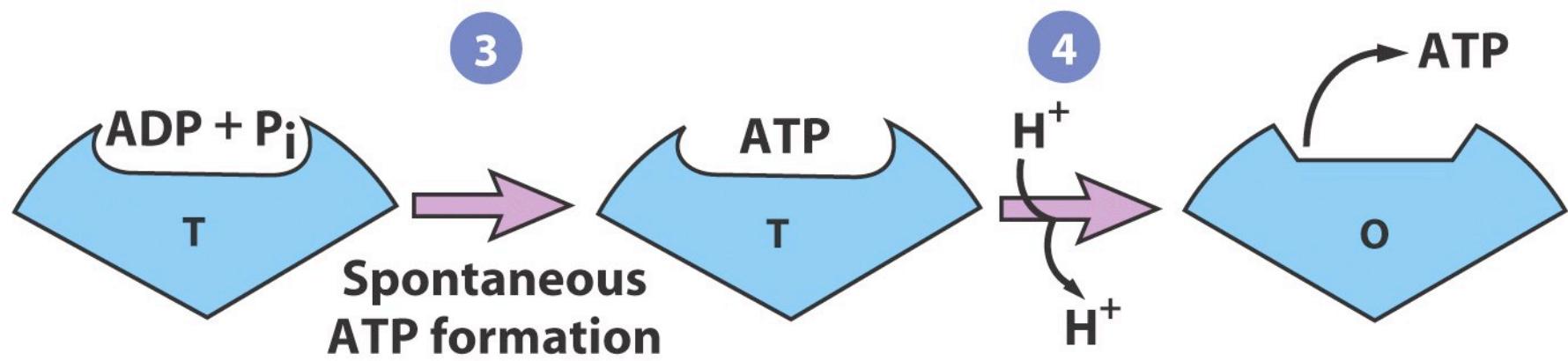


Figure 5-27a part 2 Cell and Molecular Biology, 4/e (© 2005 John Wiley & Sons)

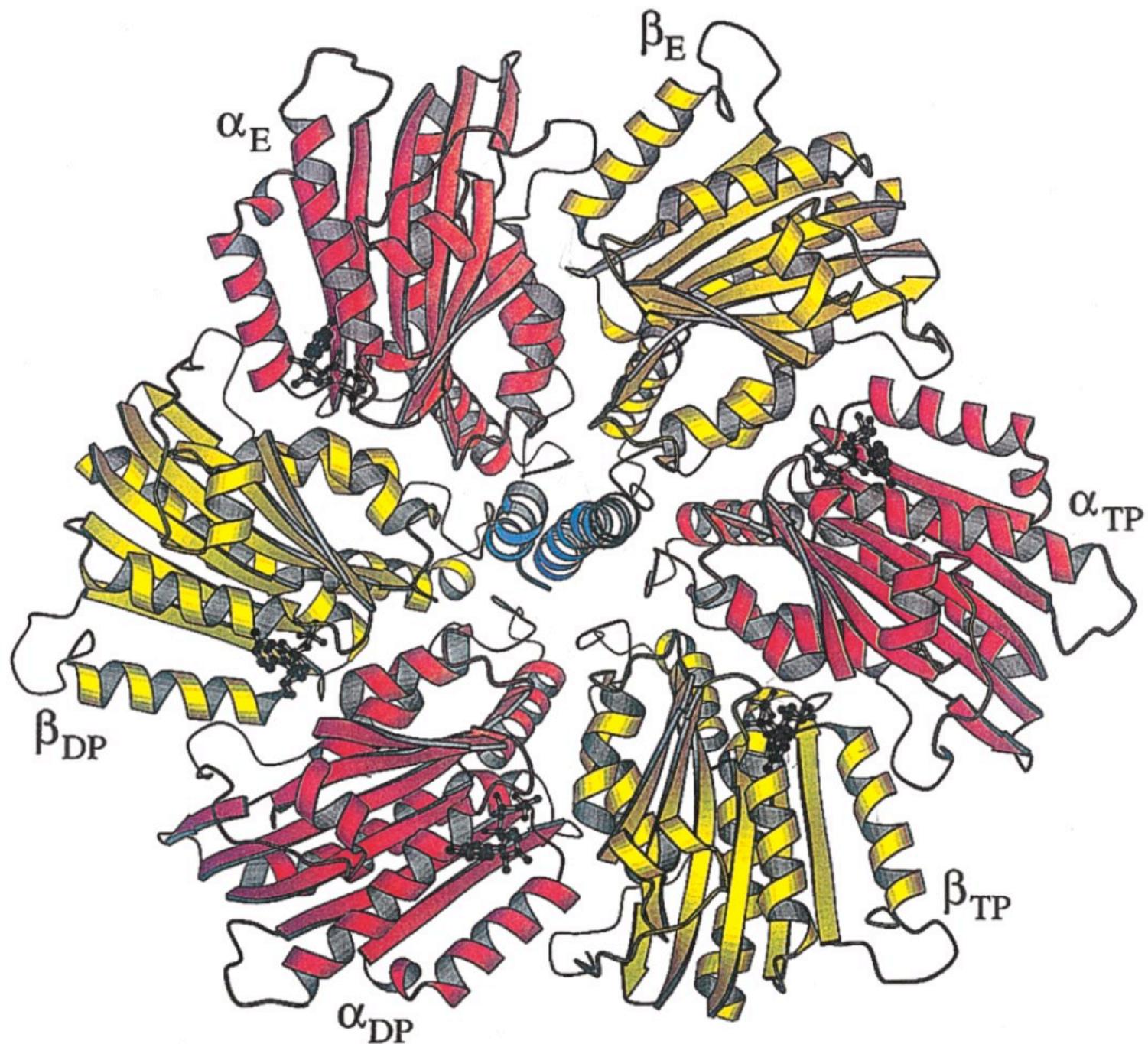


Figure 5-26b Cell and Molecular Biology, 4/e (© 2005 John Wiley & Sons)

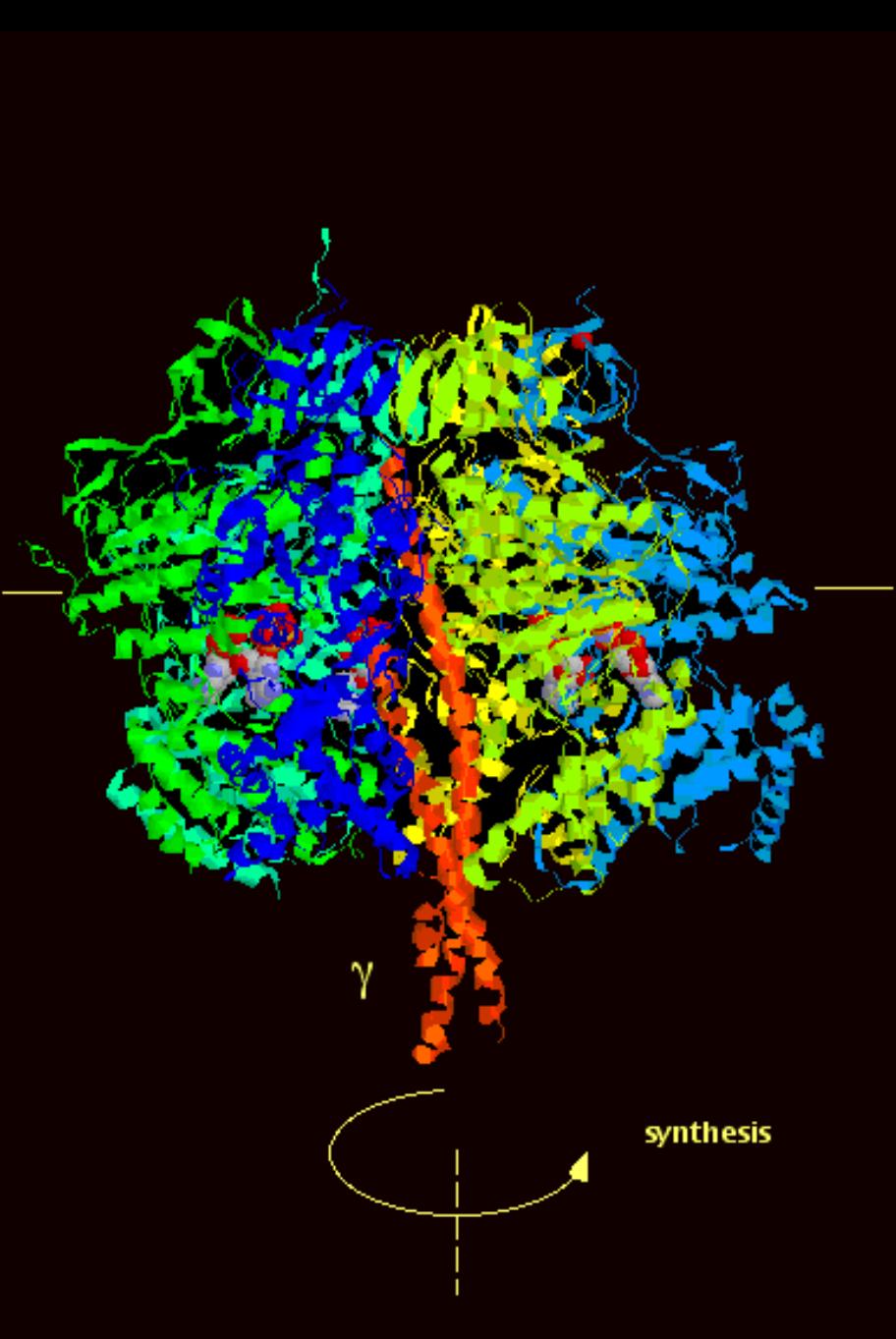


Figure from: ALLEN, J F (2002) Photosynthesis of ATP - Electrons, Proton Pumps, Rotors, and Poise. Cell 110, 273–276

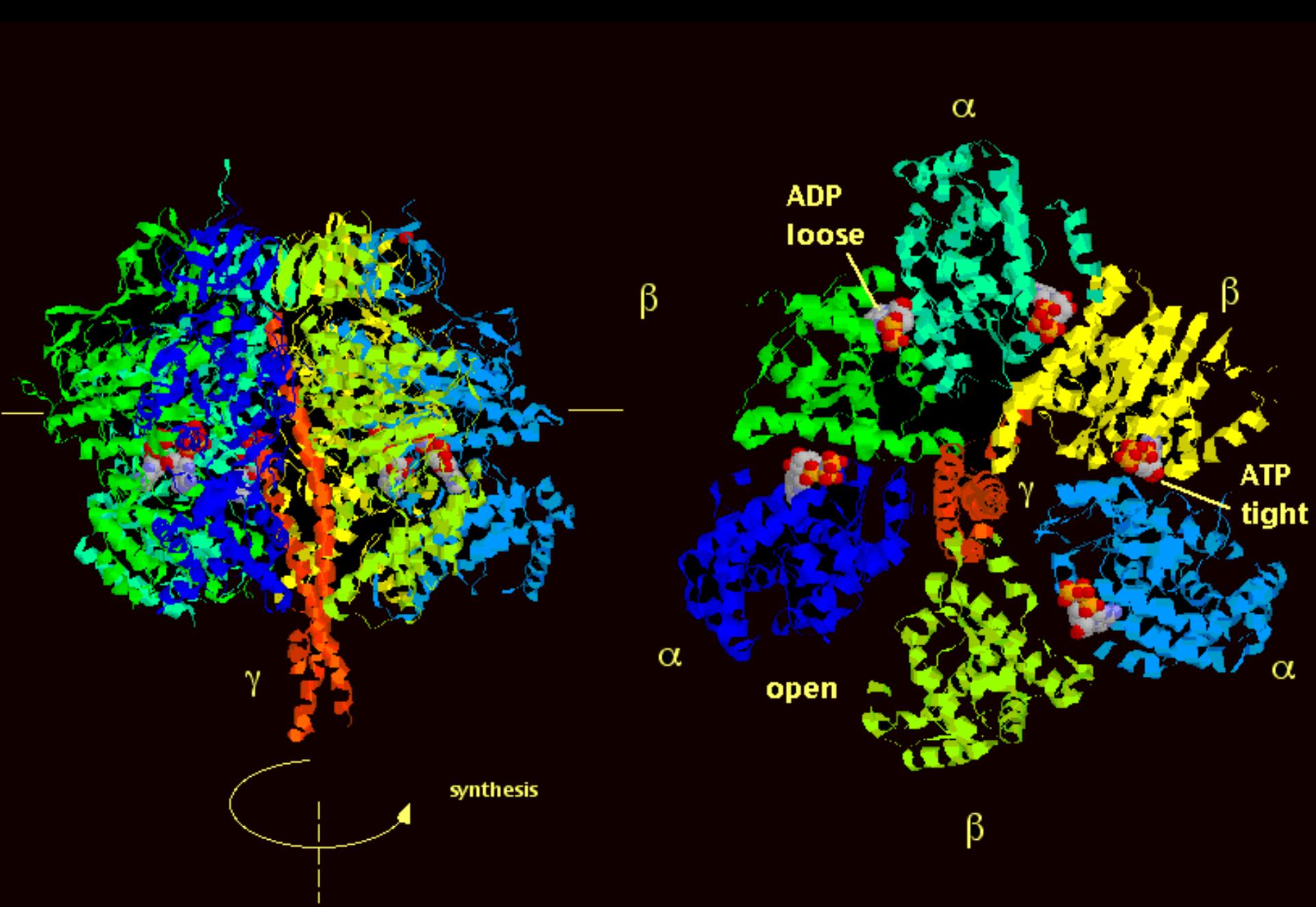


Figure from: ALLEN, J F (2002) Photosynthesis of ATP - Electrons, Proton Pumps, Rotors, and Poise. Cell 110, 273–276

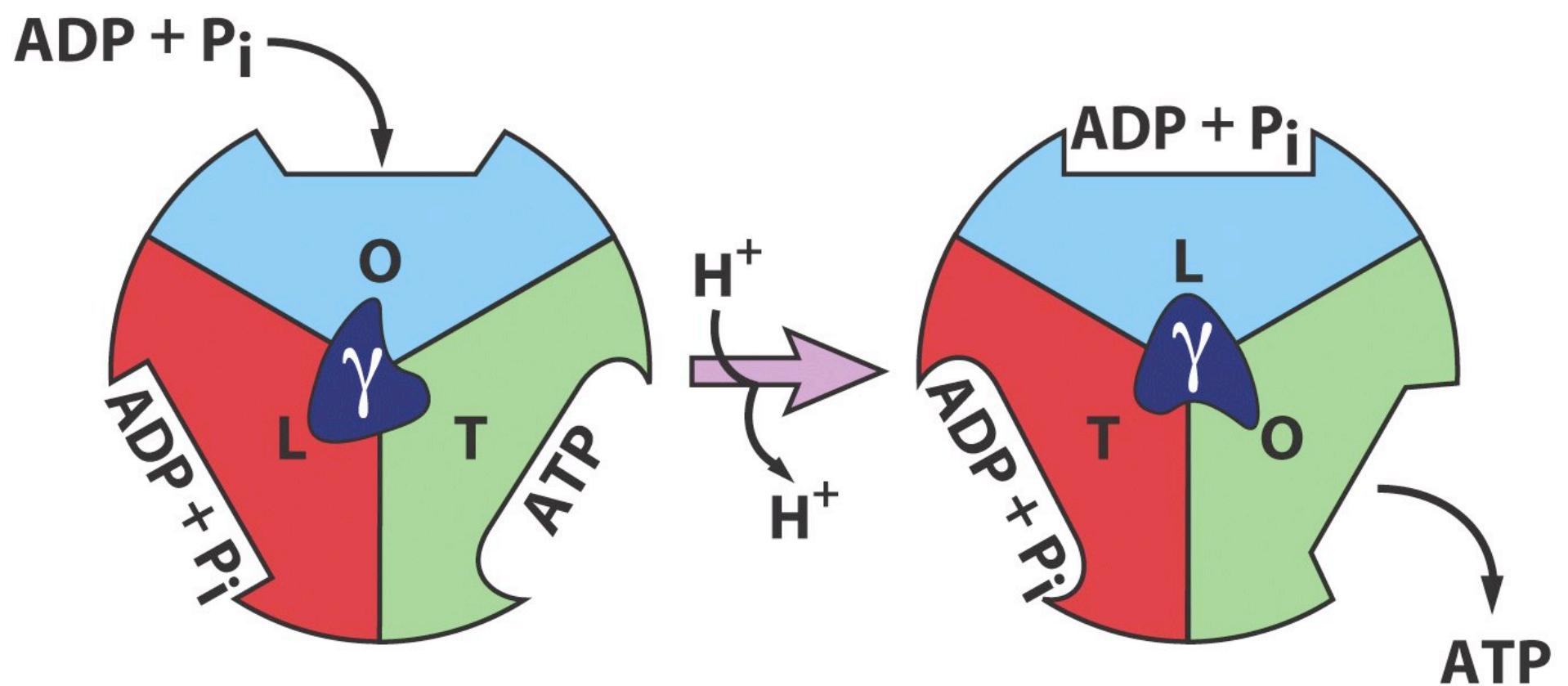


Figure 5-27b part 1 Cell and Molecular Biology, 4/e (© 2005 John Wiley & Sons)

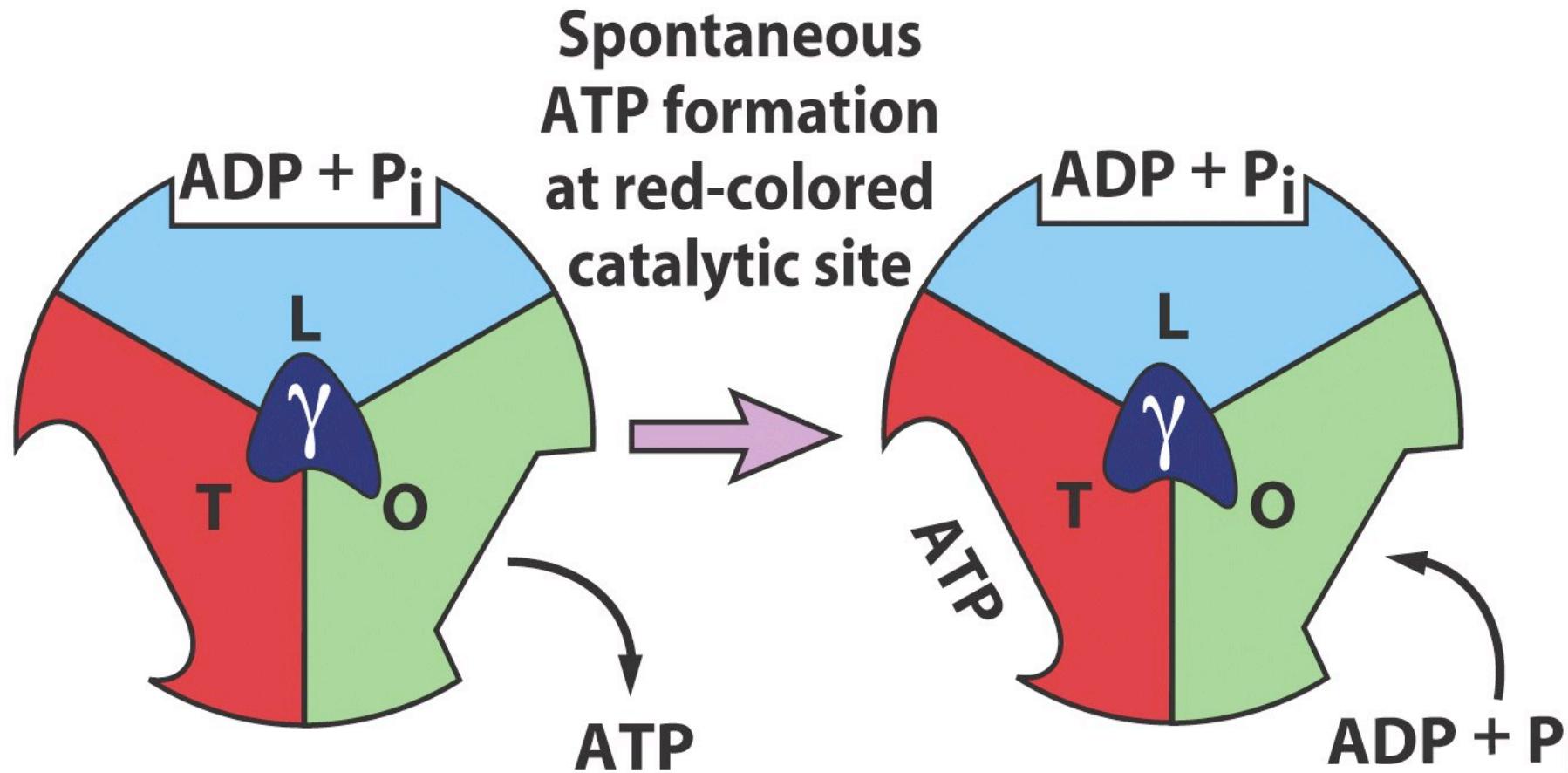


Figure 5-27b part 2 Cell and Molecular Biology, 4/e (© 2005 John Wiley & Sons)

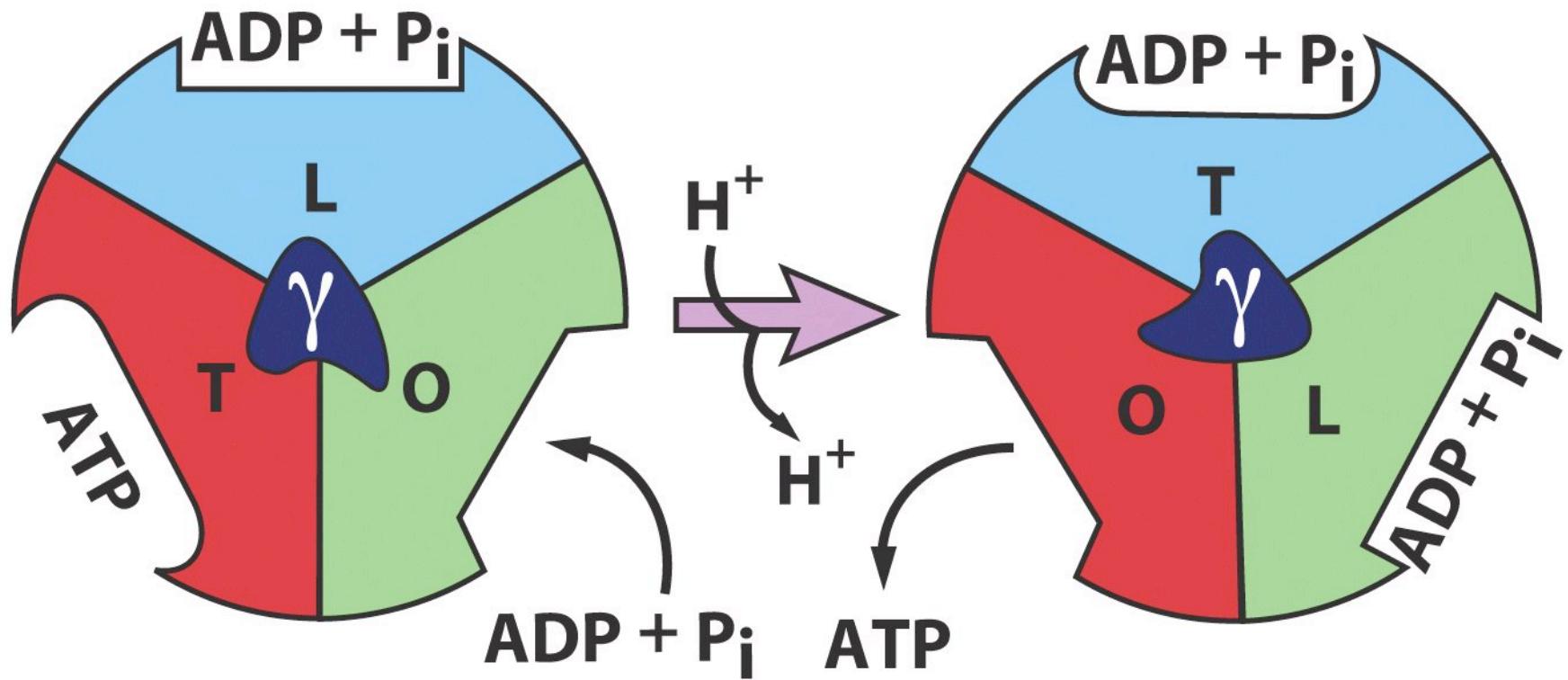


Figure 5-27b part 3 Cell and Molecular Biology, 4/e (© 2005 John Wiley & Sons)

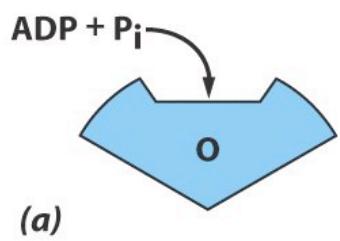


Figure 5-27 Cell and Molecular Biology, 4/e (© 2005 John Wiley & Sons)

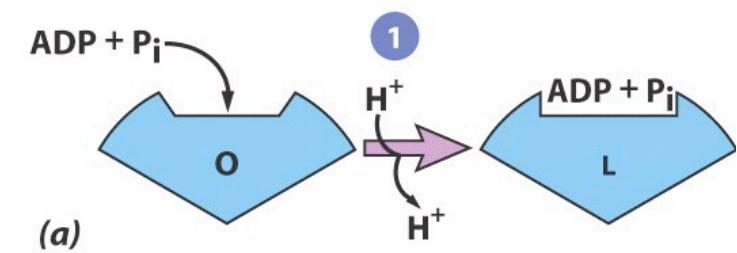


Figure 5-27 Cell and Molecular Biology, 4/e (© 2005 John Wiley & Sons)

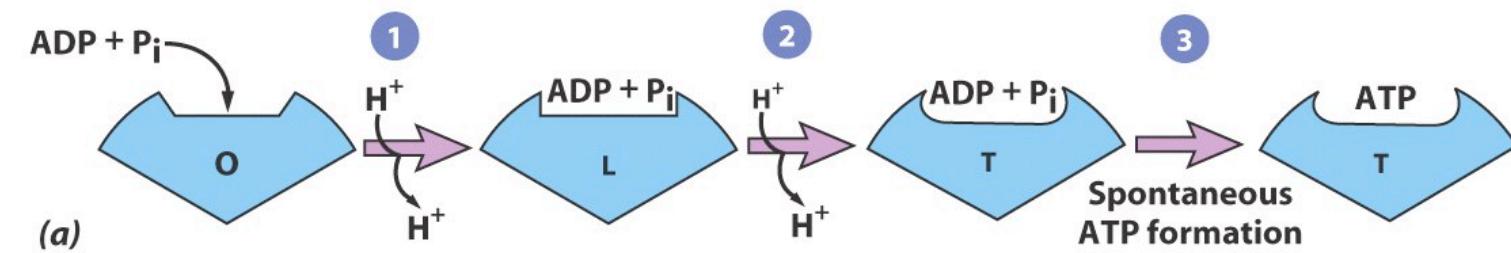


Figure 5-27 Cell and Molecular Biology, 4/e (© 2005 John Wiley & Sons)

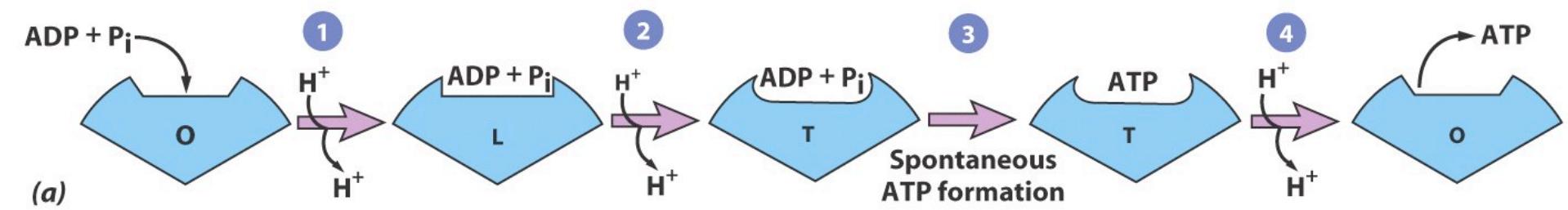


Figure 5-27 Cell and Molecular Biology, 4/e (© 2005 John Wiley & Sons)

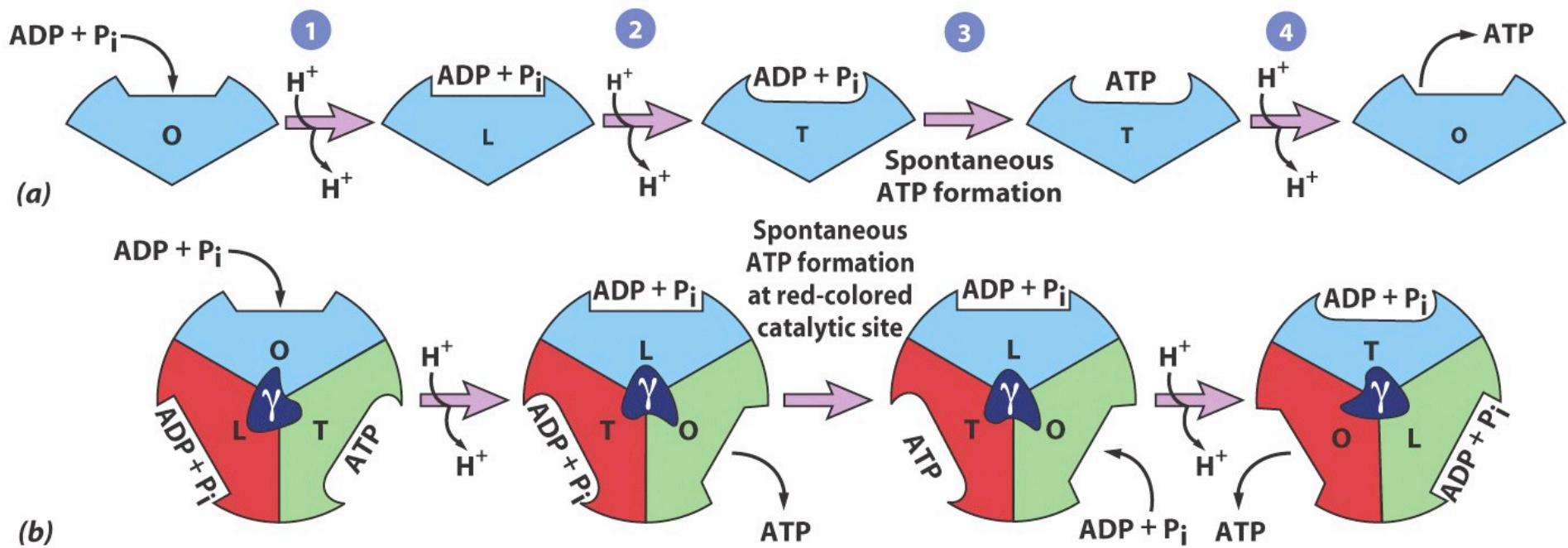
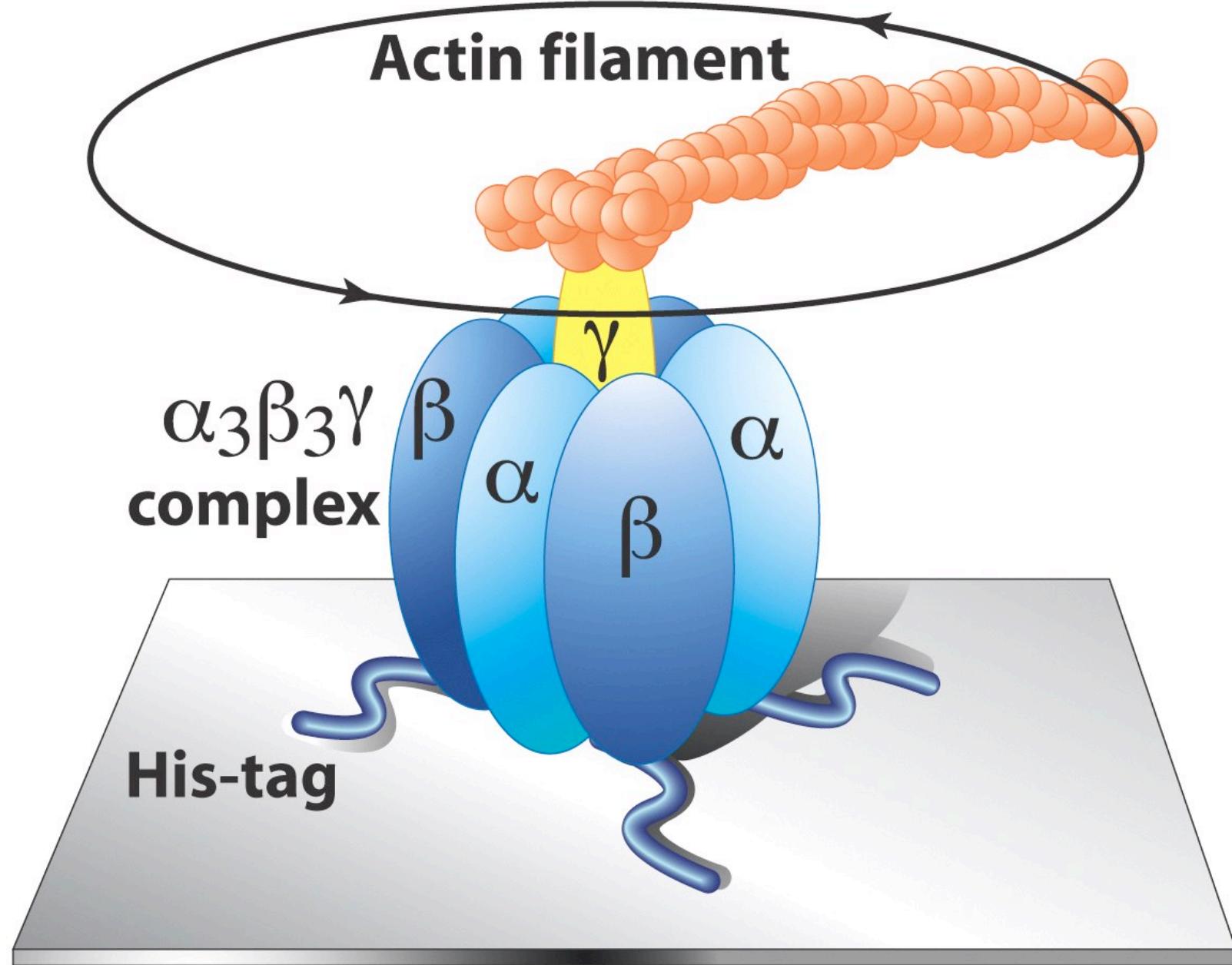


Figure 5-27 Cell and Molecular Biology, 4/e (© 2005 John Wiley & Sons)



Coverslip coated with Ni-NTA



NOBEL PRIZES

ALFRED NOBEL

PRIZE AWARDERS

NOMINATION

PRIZE ANNOUNCEMENTS

AWARD CEREMONIES

E

By Year

Nobel Prize in Physics

Nobel Prize in Chemistry

Nobel Prize in Medicine

Nobel Prize in Literature

Nobel Peace



The Nobel Prize in Chemistry 1978

"for his contribution to the understanding of biological energy transfer through the formulation of the chemiosmotic theory"

**Peter D. Mitchell**

United Kingdom

Glynn Research
Laboratories
Bodmin, United Kingdom

b. 1920

d. 1992

[Printer Friendly](#)[Comments & Questions](#)[Tell a Friend](#)**The 1978 Prize in:**

Chemistry

[Prev. year](#) [Next year](#)**The Nobel Prize in
Chemistry 1978**[Press Release](#)[Presentation Speech](#)**Peter Mitchell**[Biography](#)[Nobel Lecture](#)[Banquet Speech](#)



The Nobel Prize in Chemistry 1997

"for their elucidation of the enzymatic mechanism underlying the synthesis of adenosine triphosphate (ATP)"

**Paul D. Boyer**

1/4 of the prize

USA

University of California
Los Angeles, CA, USA

b. 1918

**John E. Walker**

1/4 of the prize

United Kingdom

MRC Laboratory of
Molecular Biology
Cambridge, United
Kingdom

b. 1941

"for the first discovery of an ion-transporting enzyme, Na^+, K^+ - ATPase"

**Jens C. Skou**

1/2 of the prize

Denmark

Aarhus University
Aarhus, Denmark

b. 1918

Printer Friendly

Comments & Questions

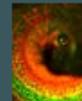
Tell a Friend

The 1997 Prize in:

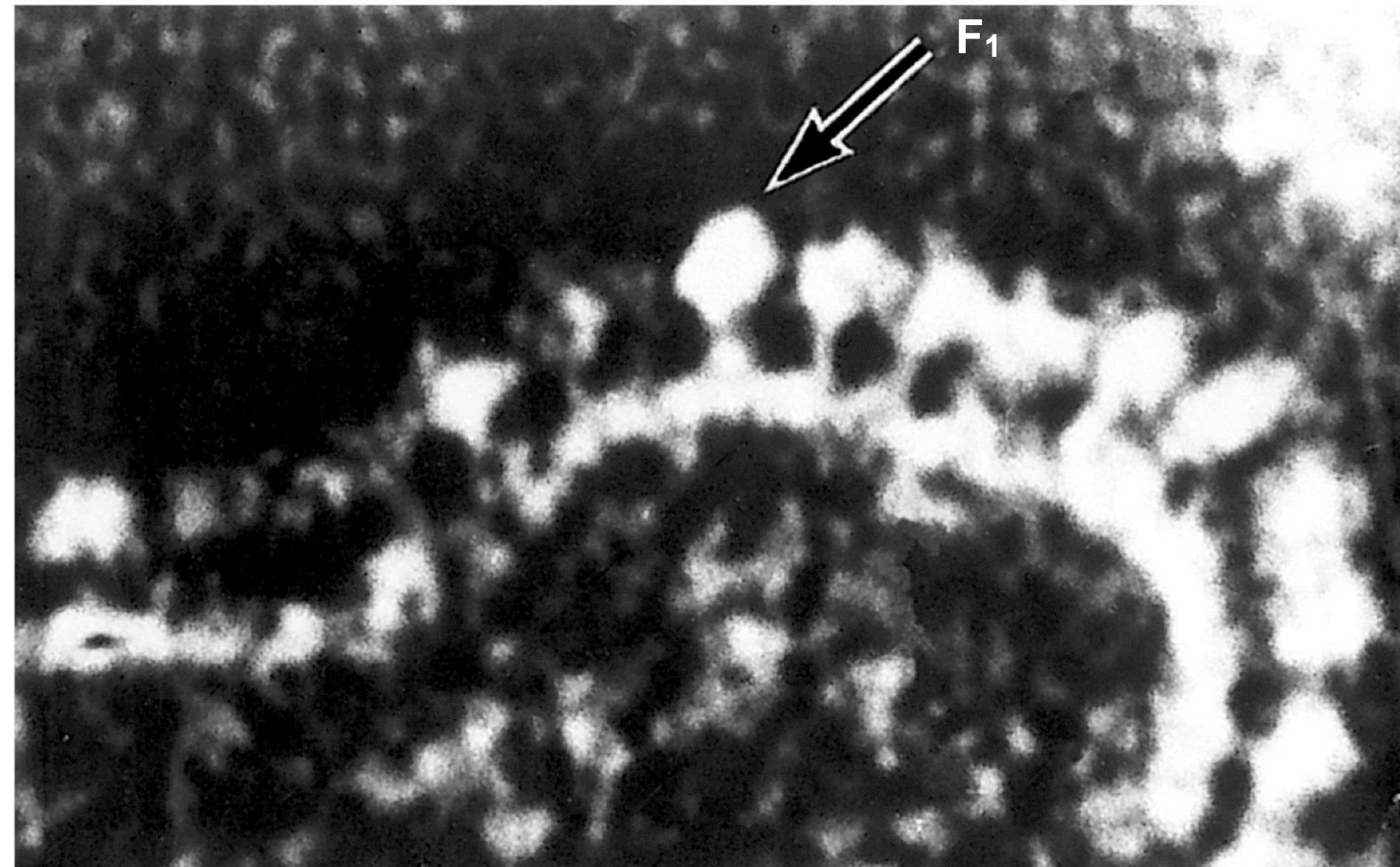
Chemistry

Prev. year

Next year

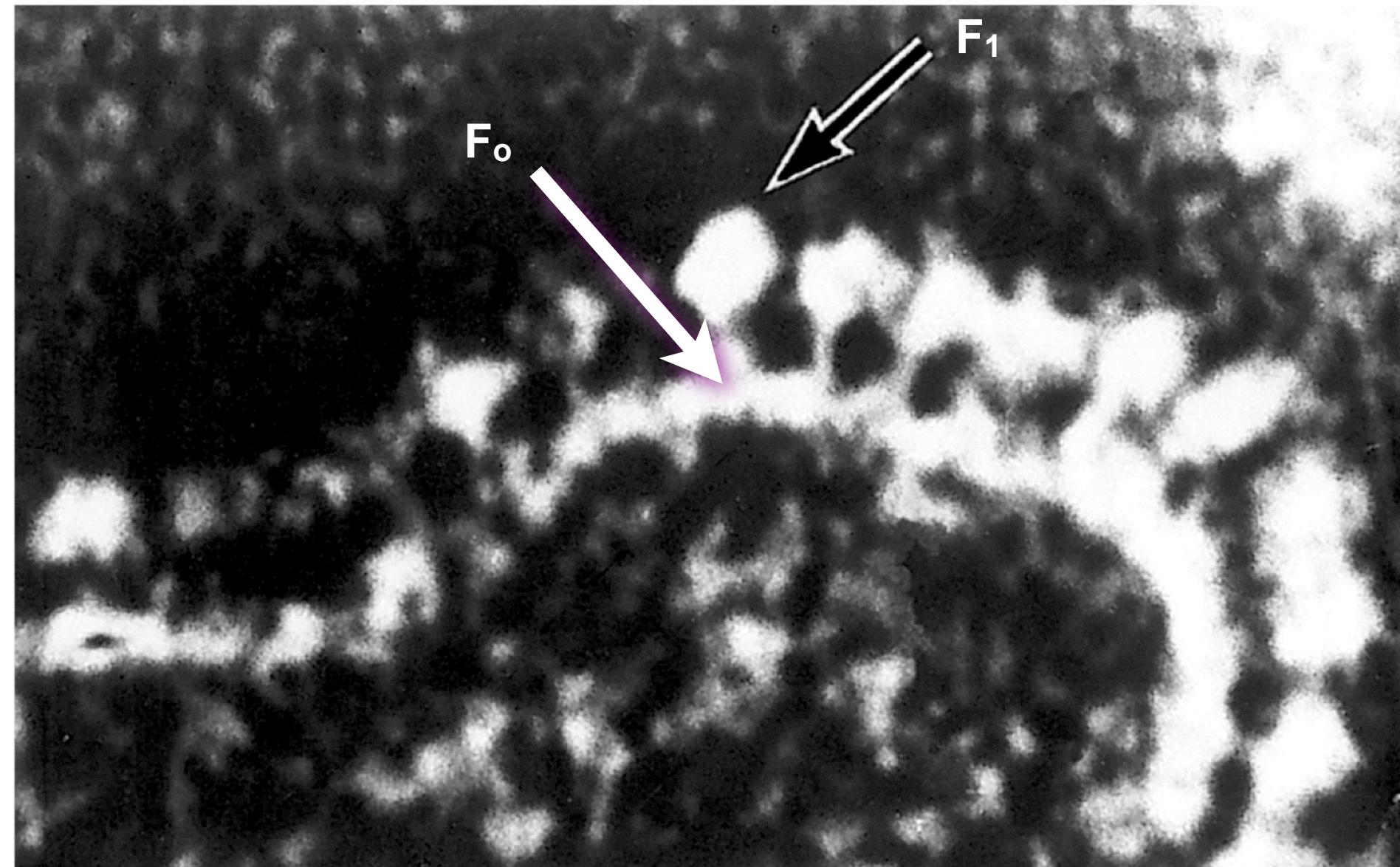
The Nobel Prize in Chemistry 1997[Press Release](#)[Presentation Speech](#)[Illustrated Presentation](#)**Paul D. Boyer**[Autobiography](#)[Nobel Lecture](#)[Interview](#)[Nobel Diploma](#)[Photo Gallery](#)[Other Resources](#)**John E. Walker**[Autobiography](#)[Nobel Lecture](#)[Nobel Diploma](#)[Photo Gallery](#)[Other Resources](#)**Jens C. Skou**[Autobiography](#)[Nobel Lecture](#)[Nobel Diploma](#)[Photo Gallery](#)[Banquet Speech](#)[Other Resources](#)**Quiz this year's Nobel Laureates****Sign up for News from Nobelprize.org****Oldest, youngest, most awarded****How do we hear? Play and learn.****Memories of that call**

F_o



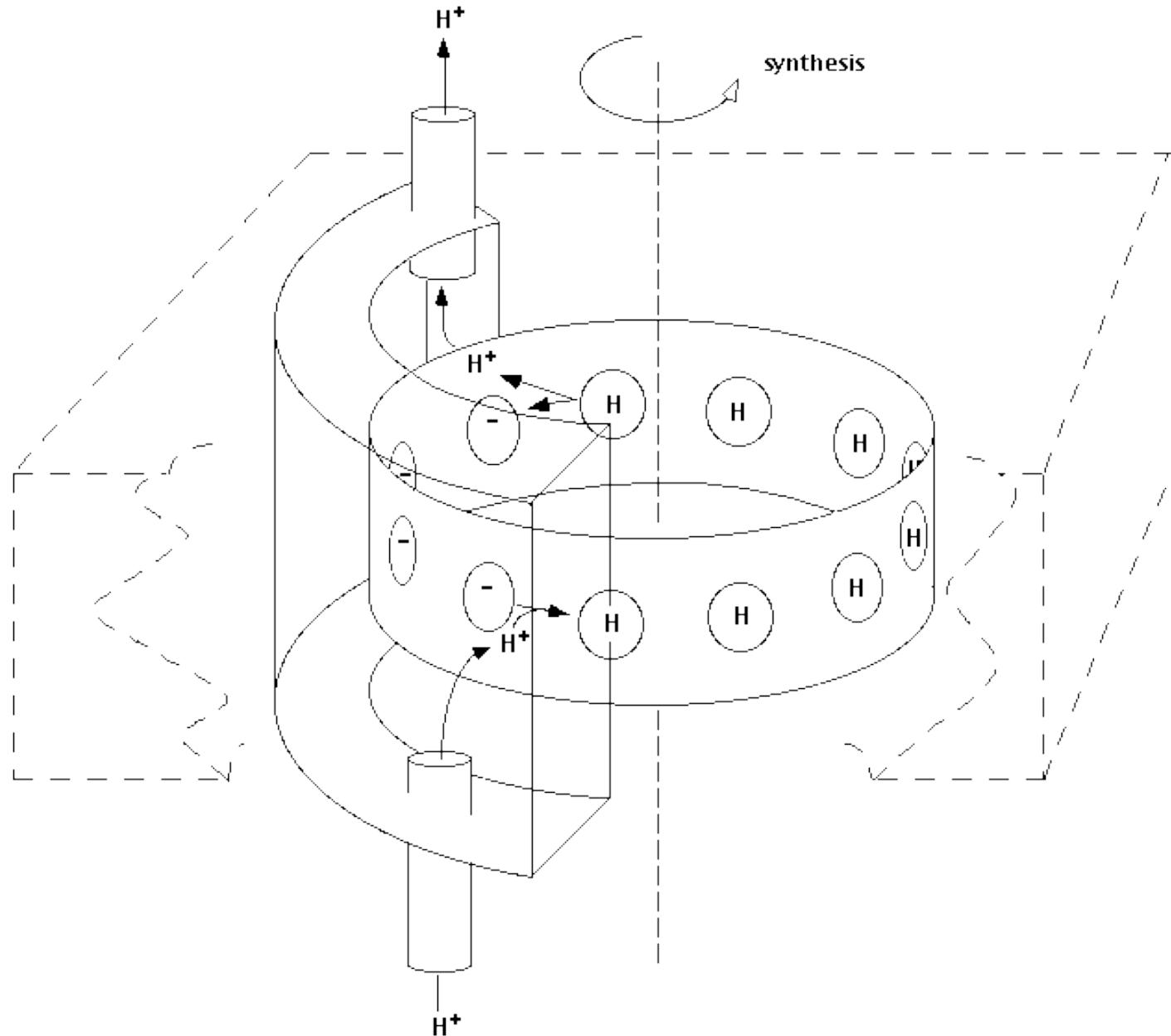
10 nm

Figure 5-21 Cell and Molecular Biology, 4/e (© 2005 John Wiley & Sons)

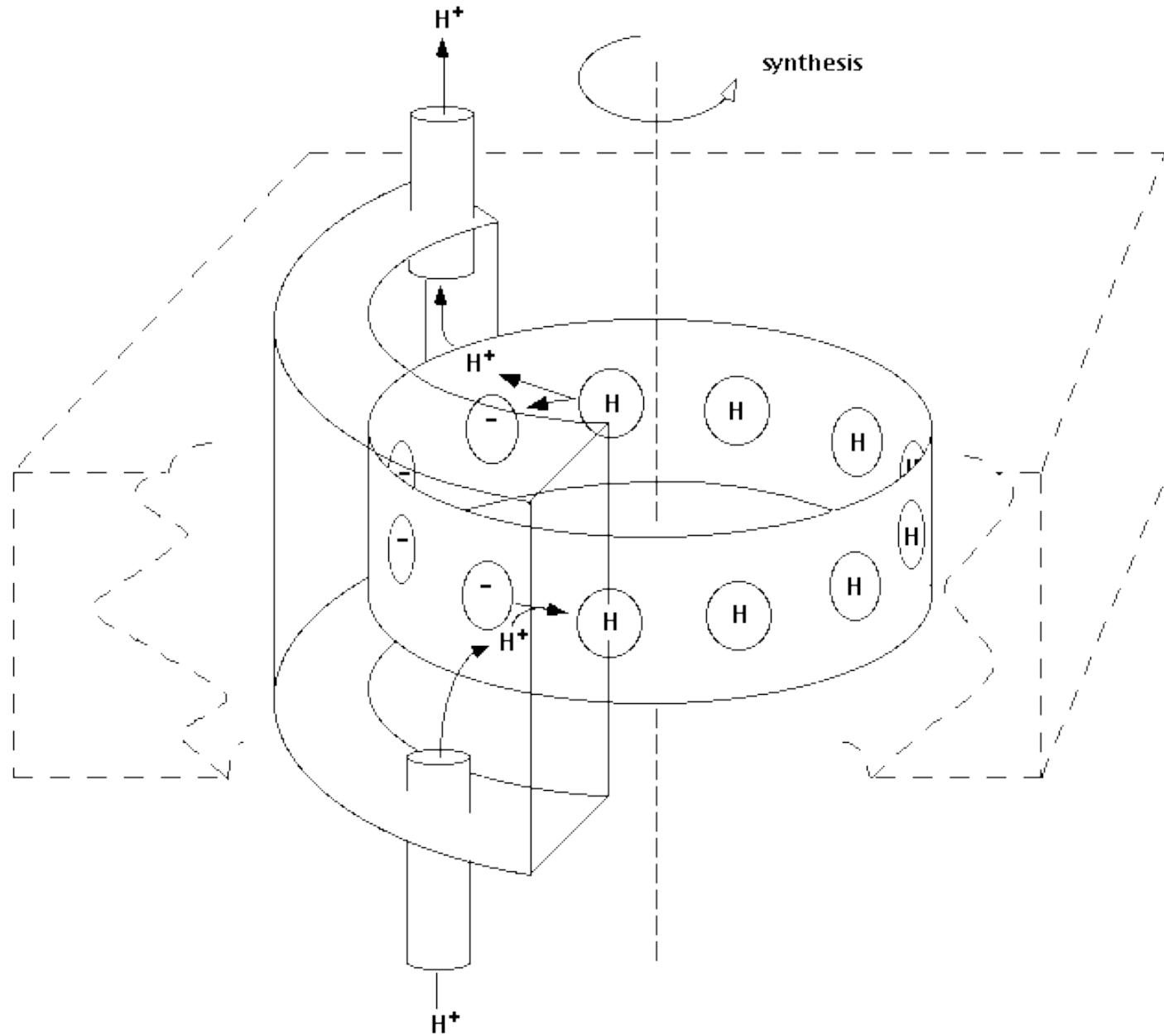


10 nm

Figure 5-21 Cell and Molecular Biology, 4/e (© 2005 John Wiley & Sons)

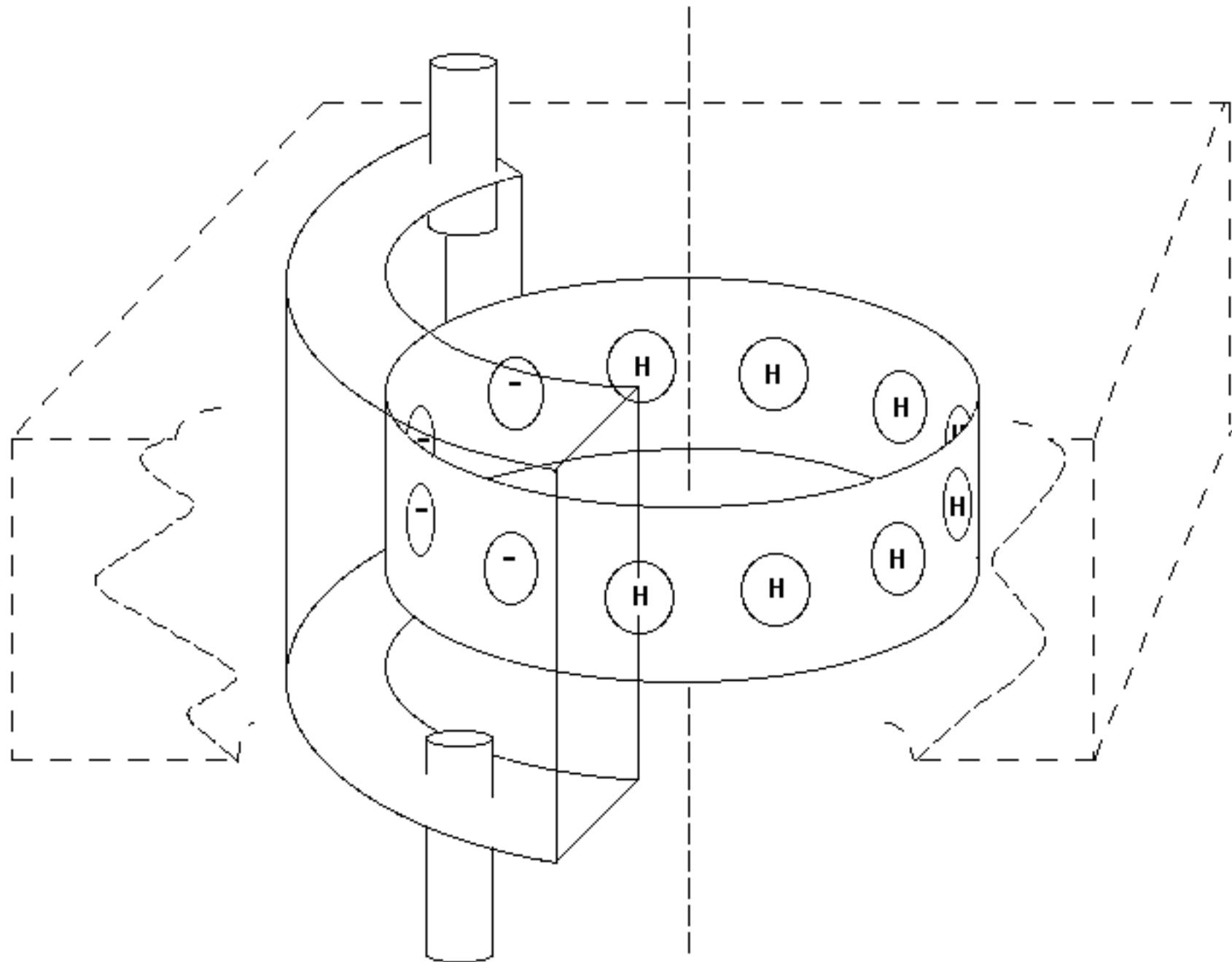


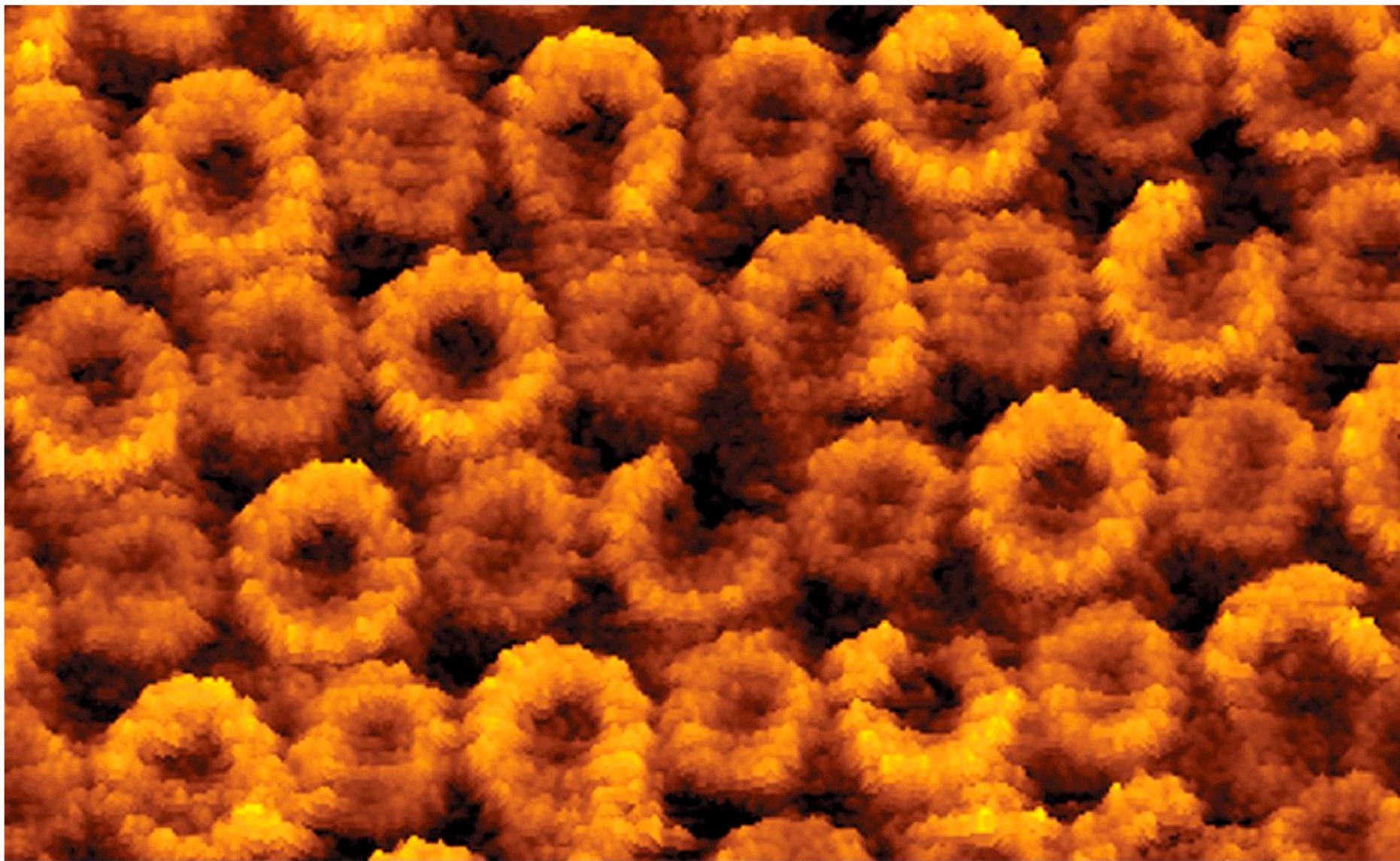
Rotation of the Fo-ATPase.Fo-ATPase as a proton-driven, rotary stepping motor, as proposed by Junge (1997).



Rotation of the Fo-ATPase. Fo-ATPase as a proton-driven, rotary stepping motor, as proposed by Junge (1997).

<http://jfa.sbccs.qmul.ac.uk/~john/webstar/ltr/06/3ATP.html>





5 nm

Figure 5-24a Cell and Molecular Biology, 4/e (© 2005 John Wiley & Sons)

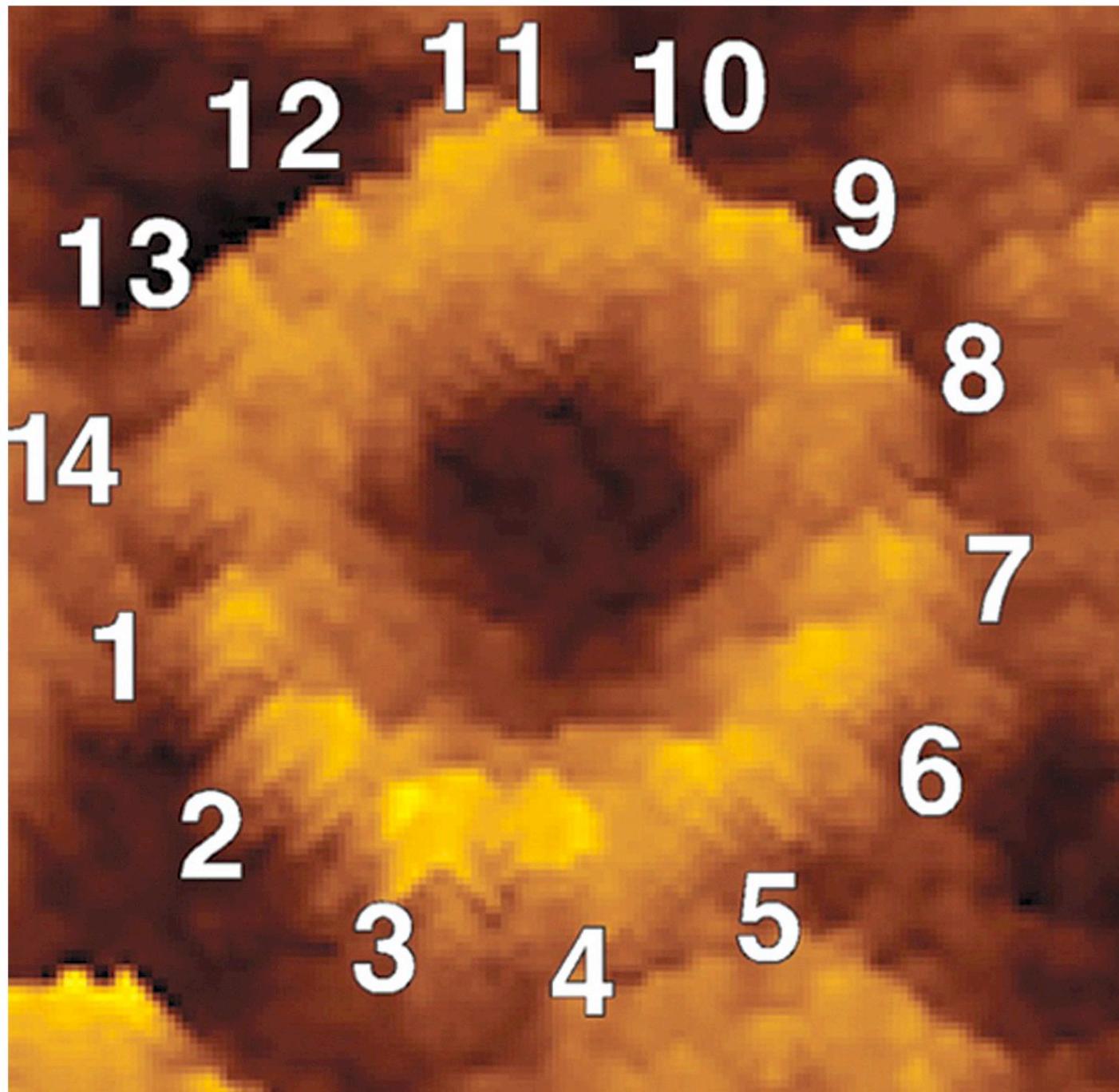


Figure 5-24b Cell and Molecular Biology, 4/e (© 2005 John Wiley & Sons)

The F₁-F₀ ATPase

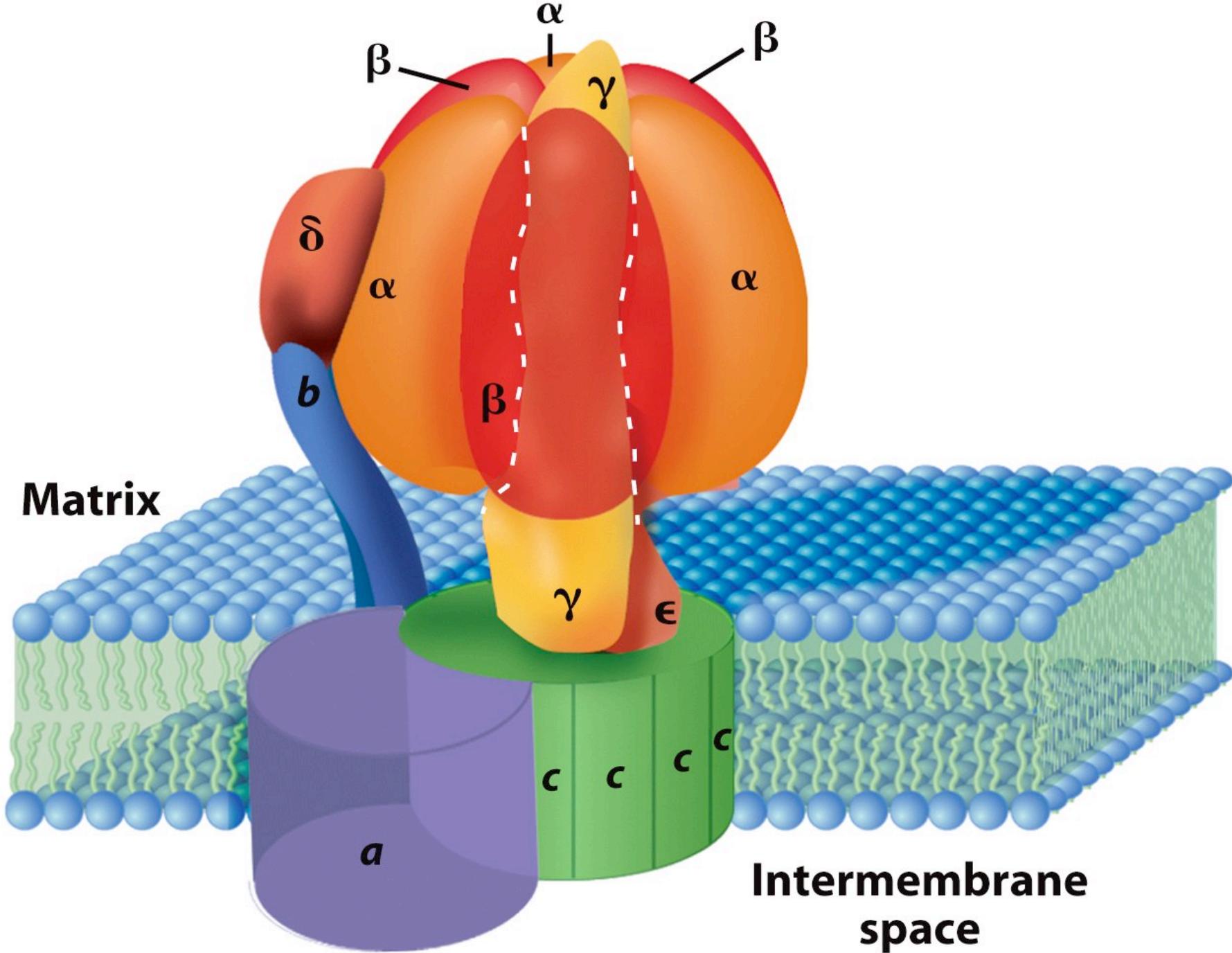


Figure 5-23b Cell and Molecular Biology, 4/e (© 2005 John Wiley & Sons)

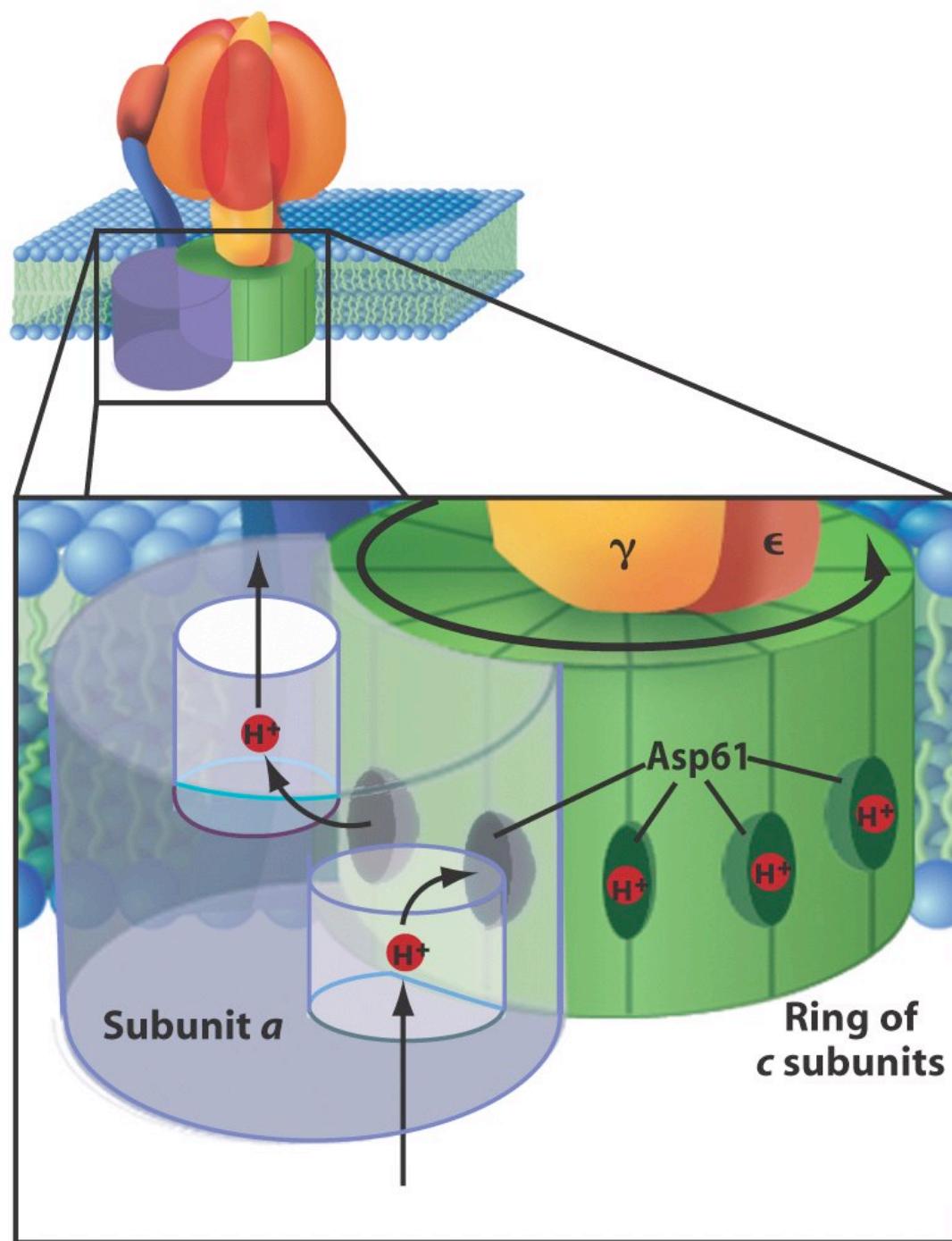


Figure 5-29 Cell and Molecular Biology, 4/e (© 2005 John Wiley & Sons)

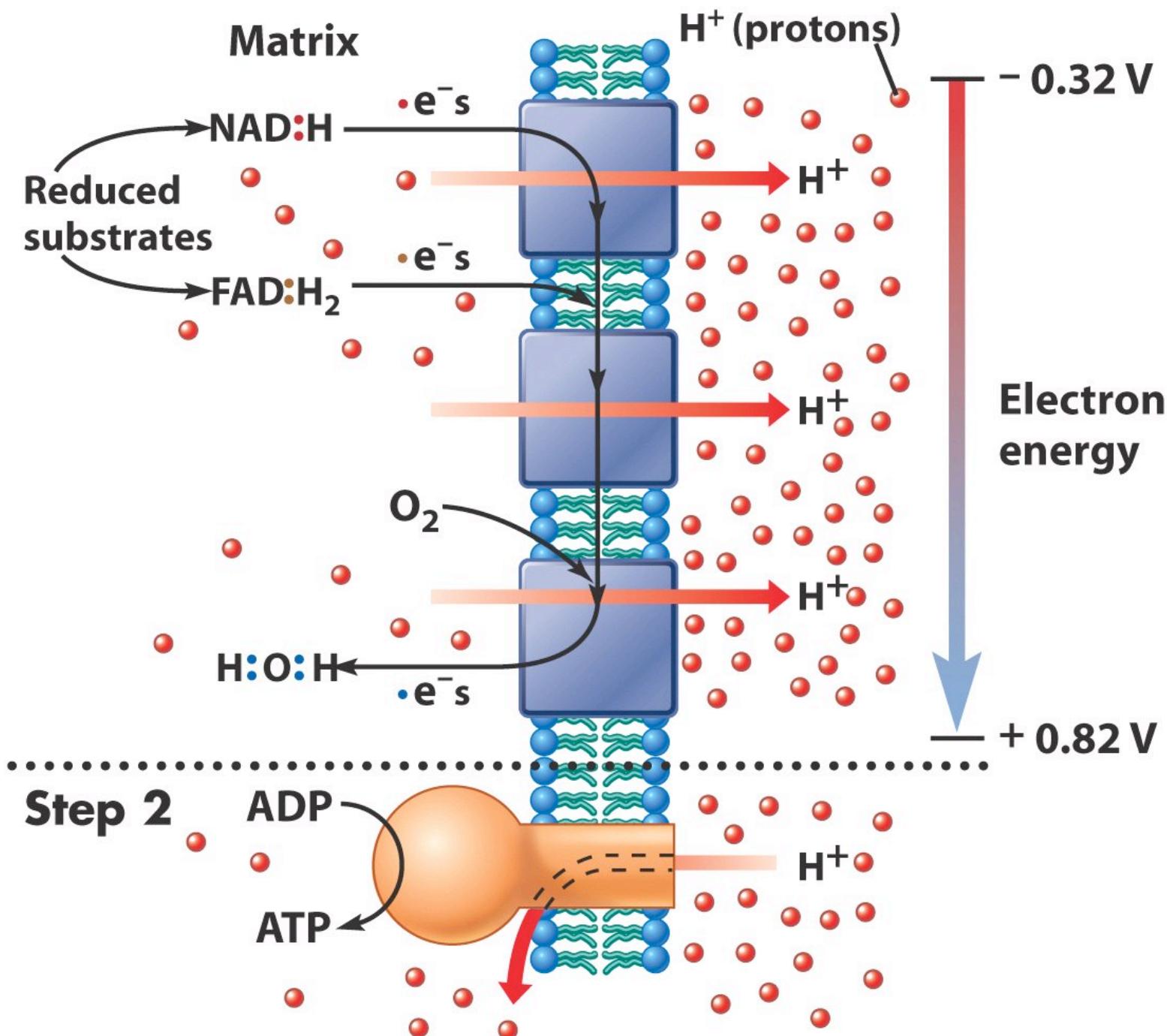


Figure 5-10 Cell and Molecular Biology, 4/e (© 2005 John Wiley & Sons)

Movies

These movies were created by Said Sannuga in collaboration with John Walker and Andrew Leslie.

Movies

These movies were created by Said Sannuga in collaboration with John Walker and Andrew Leslie.

- View from above and then below the F₁ domain along the rotating γ -subunit

Movies

These movies were created by Said Sannuga in collaboration with John Walker and Andrew Leslie.

- View from above and then below the F_1 domain along the rotating γ -subunit
- How the rotating γ -subunit imposes the conformational states on a β subunit required for substrate binding, ATP formation and ATP release

Movies

These movies were created by Said Sannuga in collaboration with John Walker and Andrew Leslie.

- View from above and then below the F_1 domain along the rotating γ -subunit
- How the rotating γ -subunit imposes the conformational states on a β subunit required for substrate binding, ATP formation and ATP release
- Three conformations of a catalytic β -subunit produced by 120° rotations of the central γ -subunit

Movies

These movies were created by Said Sannuga in collaboration with John Walker and Andrew Leslie.

- View from above and then below the F_1 domain along the rotating γ -subunit
- How the rotating γ -subunit imposes the conformational states on a β subunit required for substrate binding, ATP formation and ATP release
- Three conformations of a catalytic β -subunit produced by 120° rotations of the central γ -subunit
- Changes in the positions of sidechains in the catalytic site of F_1 -ATPase bringing about binding and subsequent hydrolysis of ATP

Movies

These movies were created by Said Sannuga in collaboration with John Walker and Andrew Leslie.

- View from above and then below the F_1 domain along the rotating γ -subunit
- How the rotating γ -subunit imposes the conformational states on a β subunit required for substrate binding, ATP formation and ATP release
- Three conformations of a catalytic β -subunit produced by 120° rotations of the central γ -subunit
- Changes in the positions of sidechains in the catalytic site of F_1 -ATPase bringing about binding and subsequent hydrolysis of ATP
- The rotary mechanism of mitochondrial ATP synthase

View from above and then below the F_1 domain
along the rotating γ -subunit

© Medical Research Council



How the rotating γ -subunit imposes conformational states on a β -subunit required for substrate binding, ATP formation and ATP release.

© Medical Research Council



Three conformations of a catalytic β -subunit produced by 120° rotations of the central γ -subunit

© Medical Research Council



Changes in the positions of side-chains in the catalytic site of F₁-ATPase bringing about binding & subsequent hydrolysis of ATP.

© Medical Research Council



The rotary catalytic mechanism of mitochondrial ATP synthase.

© Medical Research Council







Thank you for listening

Membrane Biochemistry

Lectures by

John F.Allen

School of Biological and Chemical Sciences, Queen Mary, University of London

jfallen.org/lectures

