

# Membrane Biochemistry

Lectures by  
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# Lectures in Membrane Biochemistry

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- [The endomembrane system - endocytosis and exocytosis \(Acrobat, .pdf file\)](#)
- [The endomembrane system - vesicular transport and protein trafficking \(Acrobat, .pdf file\)](#)
- [Transport across membranes 1 - Proteins \(Acrobat, .pdf file\)](#)
- [Transport across membranes 2 - Small molecules and ions \(Acrobat, .pdf file\)](#)

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Course web pages

[Membrane Biochemistry web pages](#)

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General reference

[Cell and Molecular Biology: Concepts and Experiments](#)  
Gerald Karp. Fifth Edition 2008. John Wiley & Sons Inc.

Please observe copyright on material incorporated into presentations linked from here.



## Further reading

- [Chemiosmotic coupling: The cost of living.](#) By Peter Rich. (.pdf file, 80 kb)
- [Power for Life.](#) Review of Nick Lane's book "Power Sex Suicide...." (.pdf file, 416 kb)
- [N,K-ATPase.](#) Page of Mark Hilge at Protein Biophysics, Nijmegen
- [ATP Synthase.](#) Group Pages of John Walker at the MRC Mitochondrial Biology Unit, Cambridge

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## Animations

- [The pump cycle of Na,K-ATPase.](#) By Mark Hilge at Protein Biophysics, Nijmegen
- [Animation. From Light to ATP.](#) By O. Fritzsche and W. Junge, University of Osnabrück. (.avi file, 17.7 mb)
- [Molecular animations of ATP synthase.](#) From the research group of John Walker at the MRC Mitochondrial Biology Unit, Cambridge
- [Animation. Powering the Cell: Mitochondria.](#) From BioVisions at Harvard University

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## Relevant Nobel prizes

- [1906 Nobel Prize in Physiology or Medicine to Camillo Golgi and Santiago Ramón y Cajal](#)
- [1974 Nobel Prize in Physiology or Medicine to Albert Claude, Christian de Duve and George E. Palade](#)
- [1978 Nobel Prize in Chemistry to Peter Mitchell](#)
- [1988 Nobel Prize in Chemistry to Johann Deisenhofer, Robert Huber and Hartmut Michel](#)
- [1997 Nobel Prize in Chemistry to Paul D. Boyer, John E. Walker and Jens C. Skou](#)
- [1999 Nobel Prize in Physiology or Medicine to Günter Blobel](#)



100%  
of

Membrane Biochemistry

# Oxidative phosphorylation and respiratory control

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# Photosynthetic phosphorylation in chloroplasts



## Wiley Cell and Molecular Biology

### How do plants convert light energy into ATP?

Leaves are shaped so as to optimize their exposure to sunlight.



*Click the leaf to view its cell layers.*

Step 01 of 07



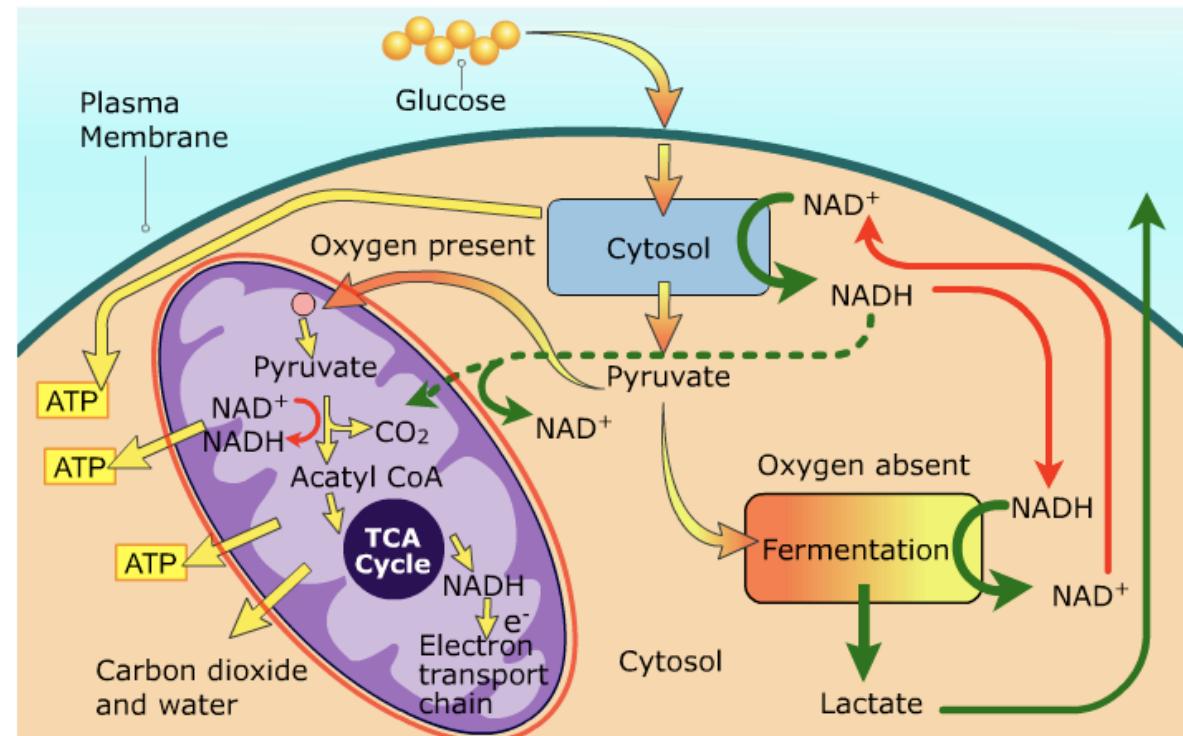
# Oxidative phosphorylation in mitochondria



## Wiley Cell and Molecular Biology

### Aerobic Respiration

Glycolysis converts glucose (C<sub>6</sub>) into two molecules of pyruvate (C<sub>3</sub>). If oxygen is present, pyruvate enters mitochondria and its free energy is utilized to make ATP via the TCA cycle and oxidative phosphorylation.



Click the mitochondrion to examine the TCA cycle and oxidative phosphorylation.

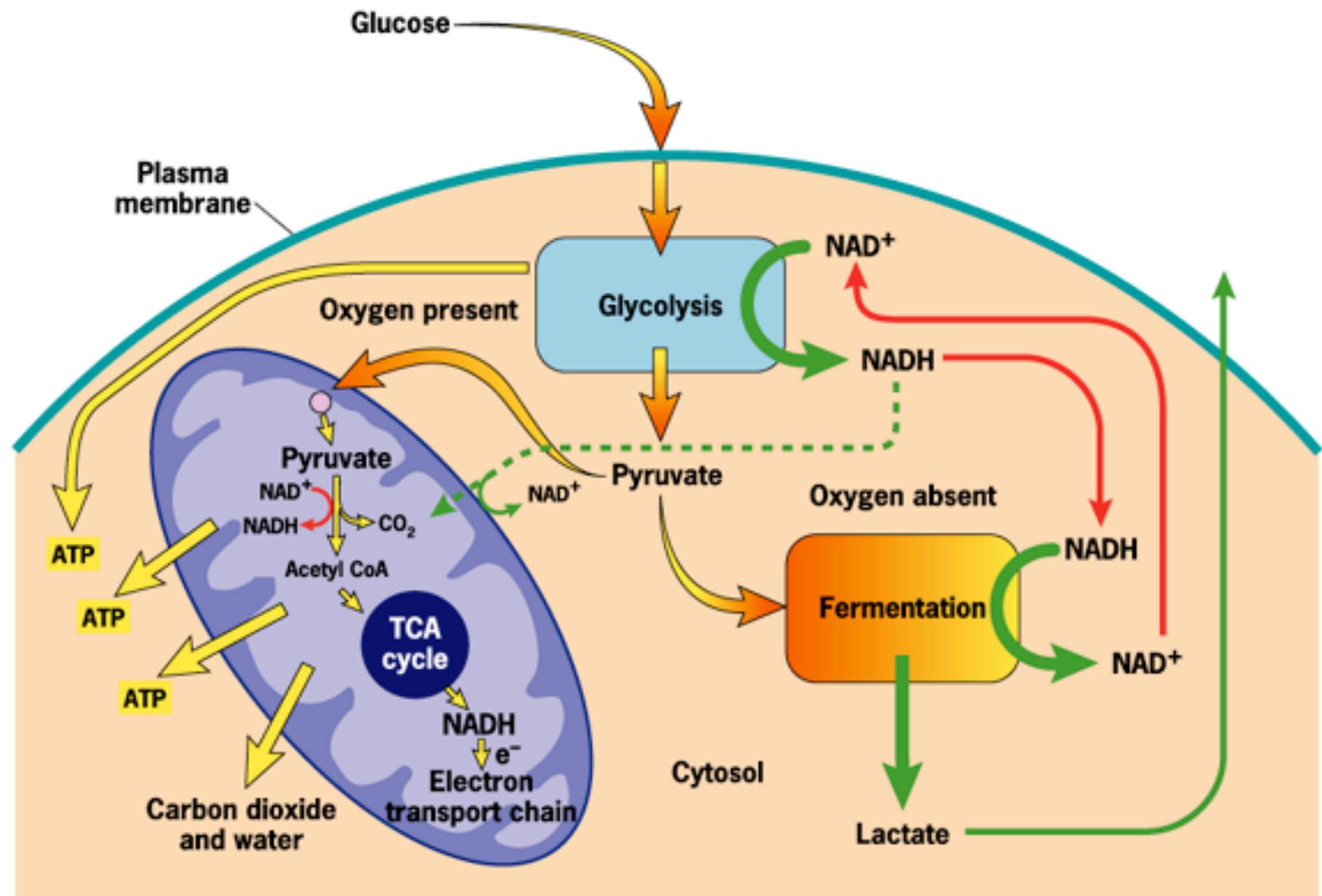


Figure 5.5 An overview of carbohydrate metabolism in eukaryotic cells.

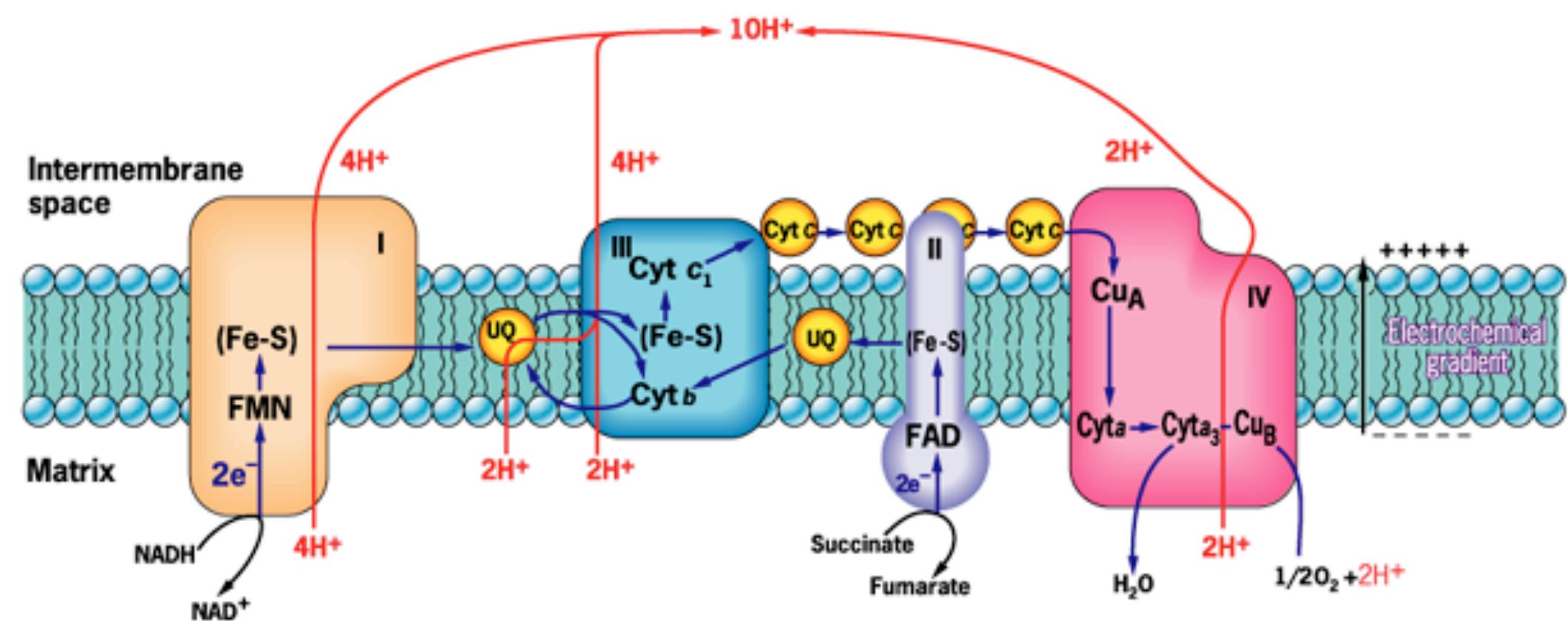


Figure 5.16 Schematic diagram of the components of the electron-transport chain within the inner mitochondrial membrane.

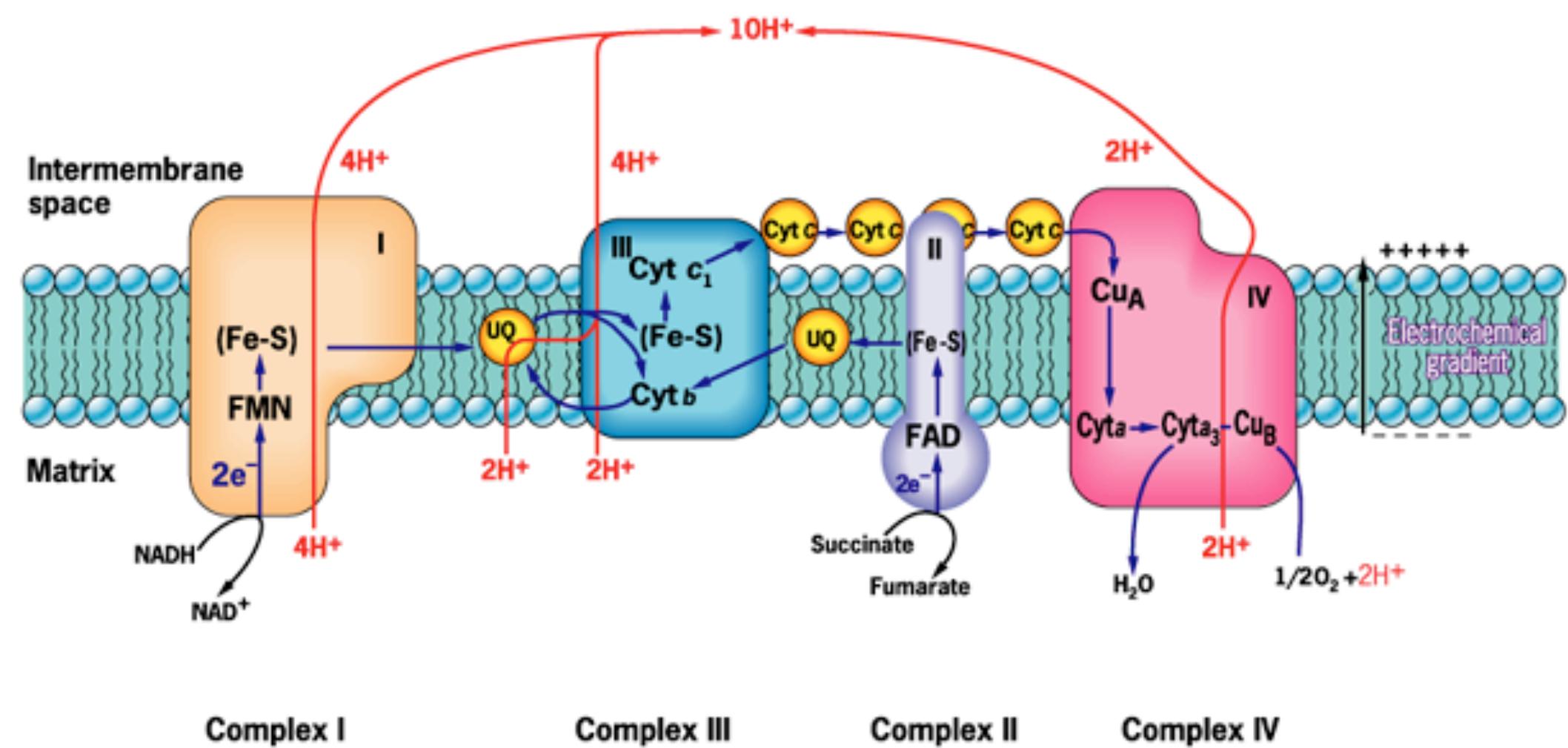


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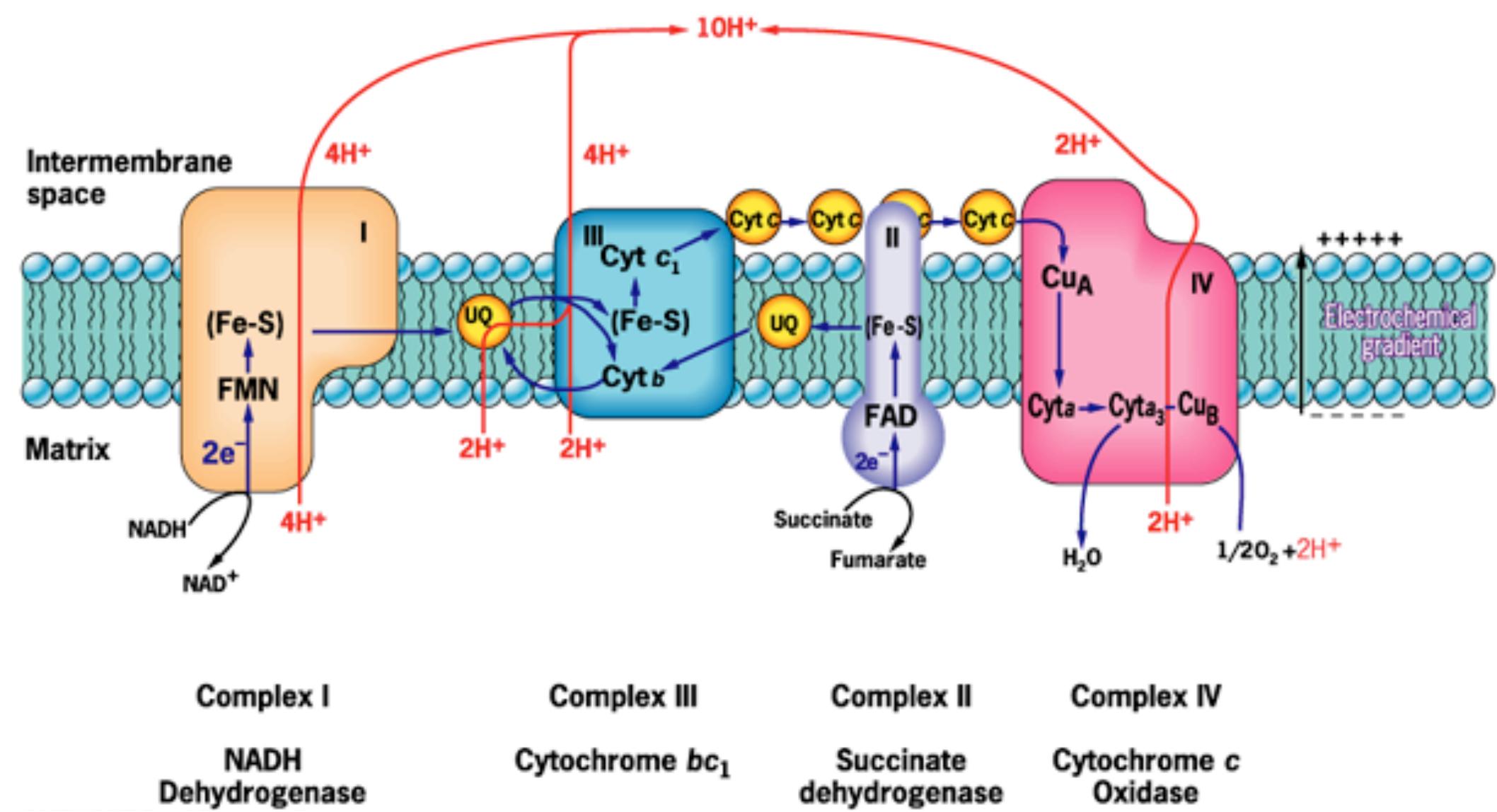


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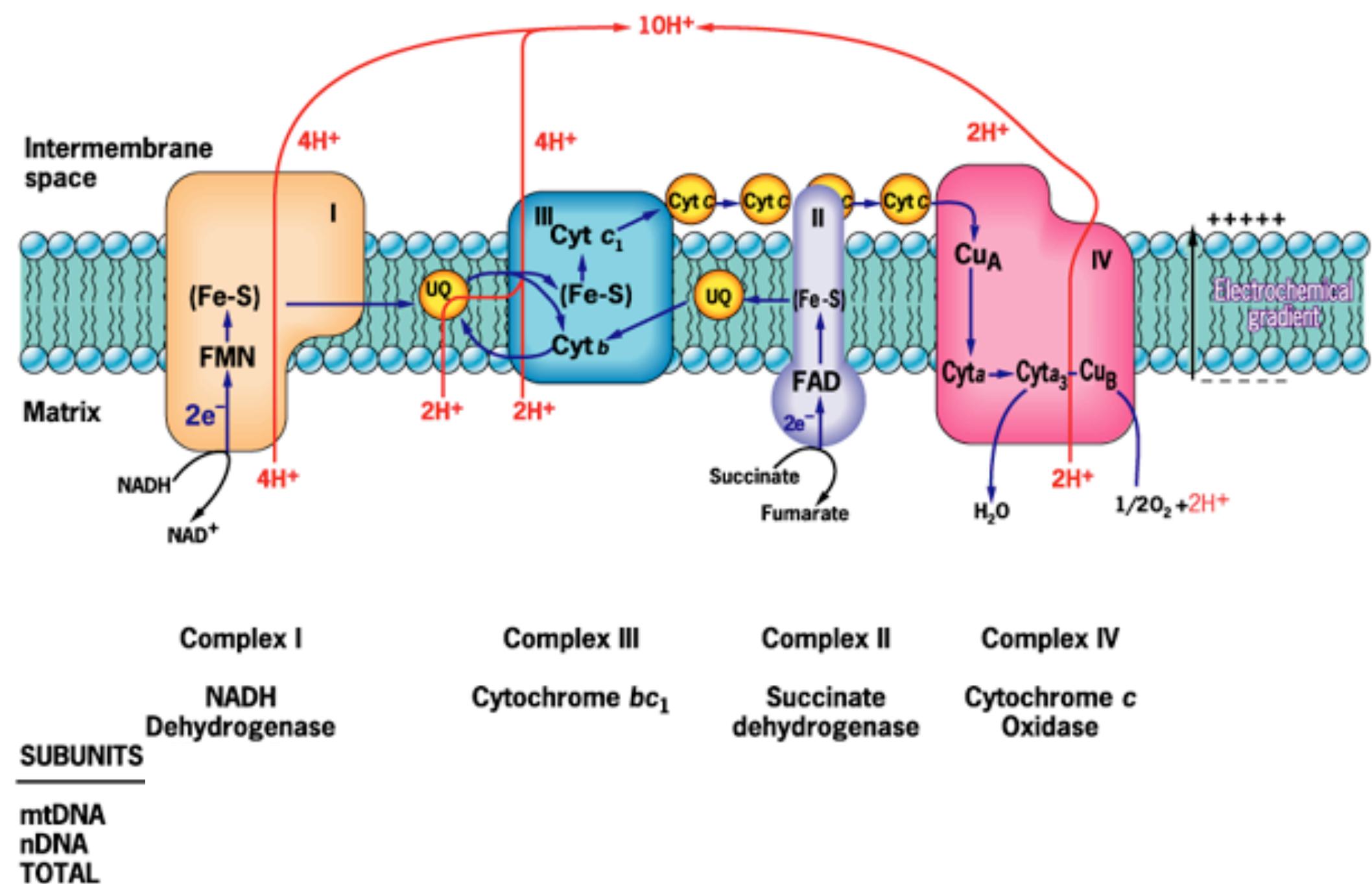
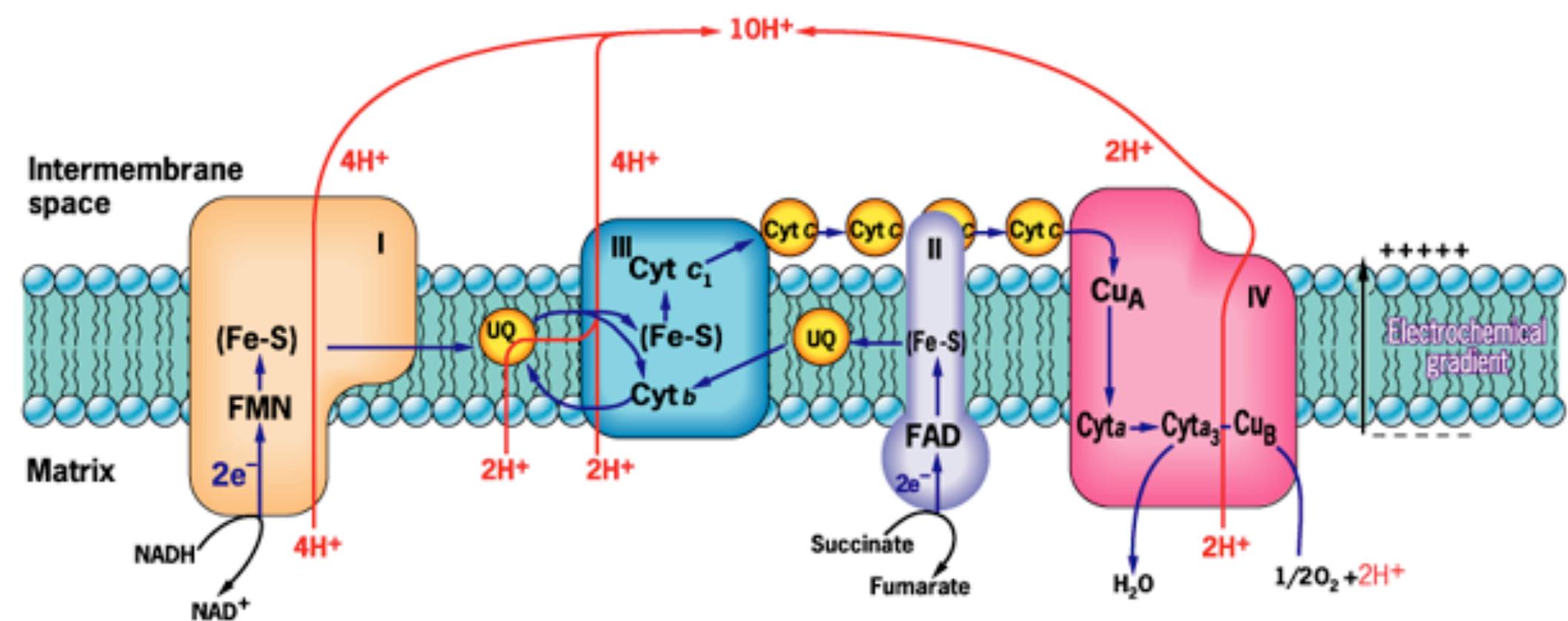
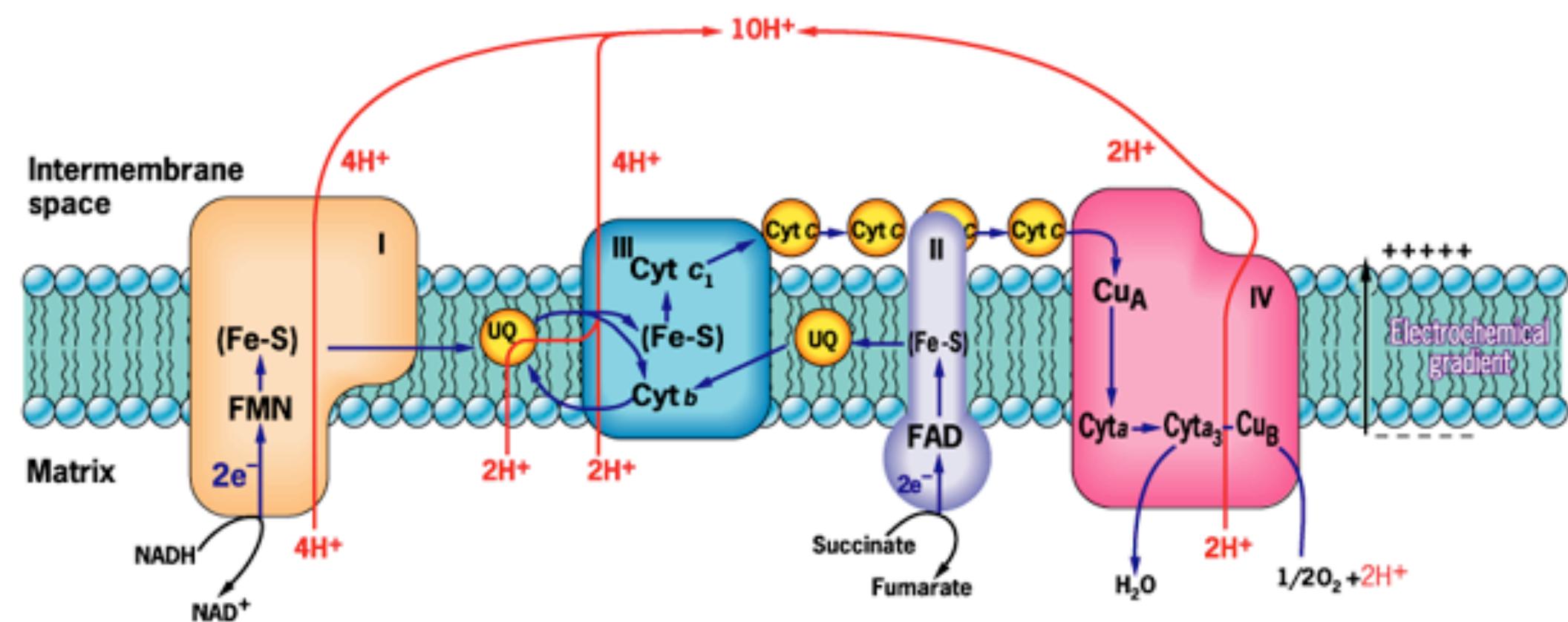


Figure 5.16 Schematic diagram of the components of the electron-transport chain within the inner mitochondrial membrane.



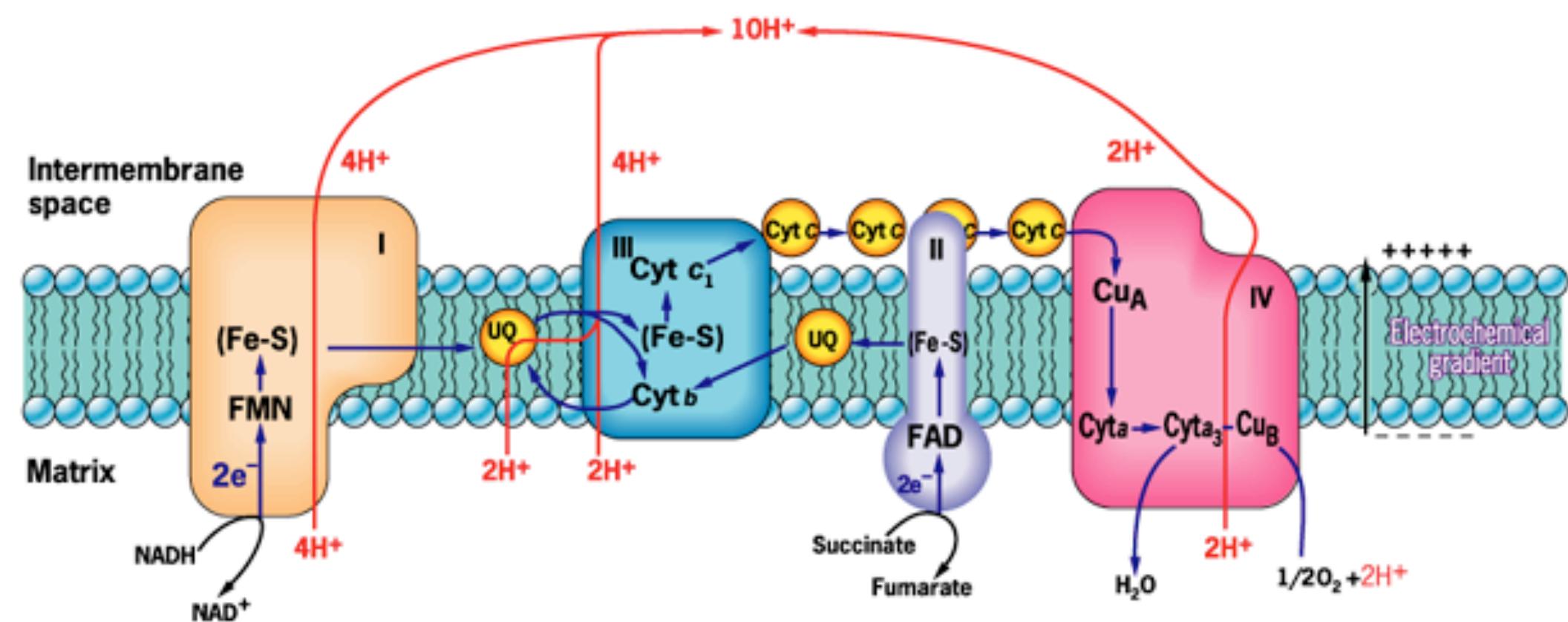
	Complex I	Complex III	Complex II	Complex IV
SUBUNITS	NADH Dehydrogenase	Cytochrome <i>bc</i> <sub>1</sub>	Succinate dehydrogenase	Cytochrome c Oxidase
mtDNA	7			
nDNA	35			
TOTAL	42			

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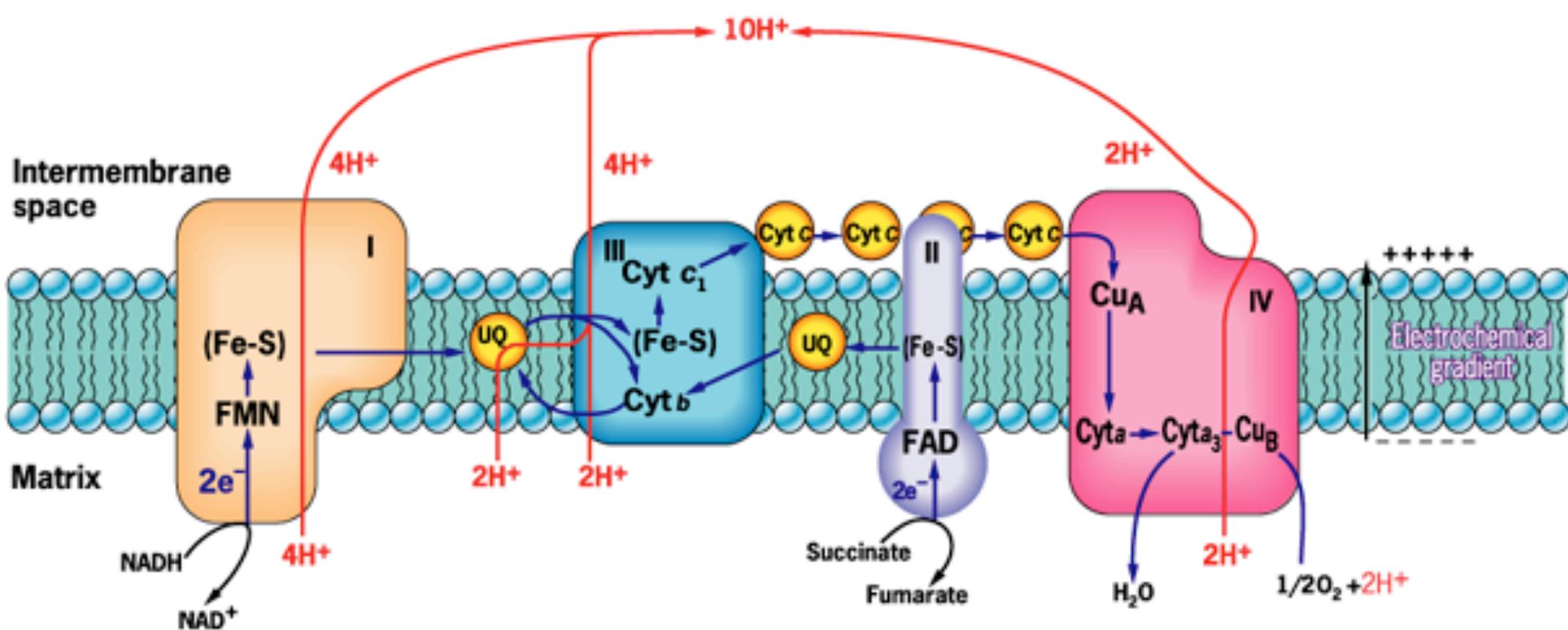
SUBUNITS	Complex I	Complex III	Complex II	Complex IV
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TOTAL	42	11		

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	Complex I	Complex III	Complex II	Complex IV
SUBUNITS	NADH Dehydrogenase Mammalian	Cytochrome <i>bc</i> <sub>1</sub>	Succinate dehydrogenase	Cytochrome c Oxidase
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## Characteristics of oxidative phosphorylation

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- a) the rate of electron transport – the number of electrons transferred along the chain in unit time – by measuring the rate at which oxygen is consumed (1 0 atom =  $2e^-$ )
- b) the amount of ATP synthesised (or amount of ADP or Pi converted into ATP)

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You will observe 1-4 in the laboratory classes.

## Characteristics of oxidative phosphorylation

- 1] Energy coupling sites**
- 2] Respiratory control**
- 3] Uncoupling agents**
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# 1] Energy Coupling Sites

It became apparent that the amount of ATP formed (or ADP/Pi converted into ATP) was stoichiometric with amount of oxygen consumed

This stoichiometry is called

P/O or  $P/2e^-$  quotient

ATP/O or  $ATP/2e^-$  quotient

ADP/O or  $ADP/2e^-$  quotient

The value obtained reflects several parameters

- 1) the nature of the substrate undergoing oxidation
- 2) the integrity of coupling membrane
- 3) the redox carrier composition of respiratory chain



In late 1940s and early 1950s several labs showed that oxidation of NADH (in fact oxidation of substrates like  $\beta$ HOB that feed into resp. chain via NADH) leads to production of 3 molecules of ATP for every oxygen atom consumed, that is, for every pair of electrons ( $2e^-$ ) transferred through the chain (ATP/O quotient of 3).

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It is possible to isolate particular sections of the respiratory chain with different electron donors and acceptors, together with electron transfer inhibitors to inhibit other parts of the chain.

<b>Reducant</b>	<b>Oxidant</b>	<b>Inhibitor</b>	<b>ATP/O</b> <b>ATP/2e<sup>-</sup></b>

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An increased ATP/2e<sup>-</sup> quotient is observed if it is measured with ADP and Pi already inside (endogenous) mitochondria. About 25% of energy released by electron transfer is used for this active transport of ATP, ADP and Pi.

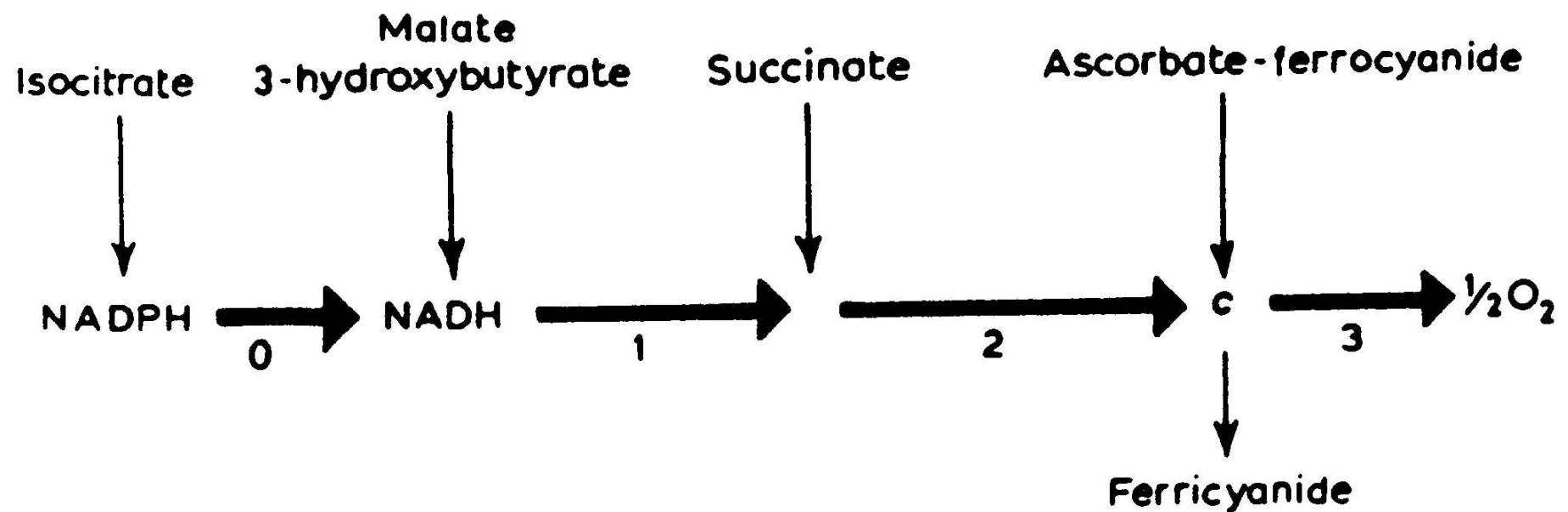


Fig. 4.1 Energy coupling sites.

## Characteristics of oxidative phosphorylation

- 1] Energy coupling sites
- 2] **Respiratory control**
- 3] Uncoupling agents
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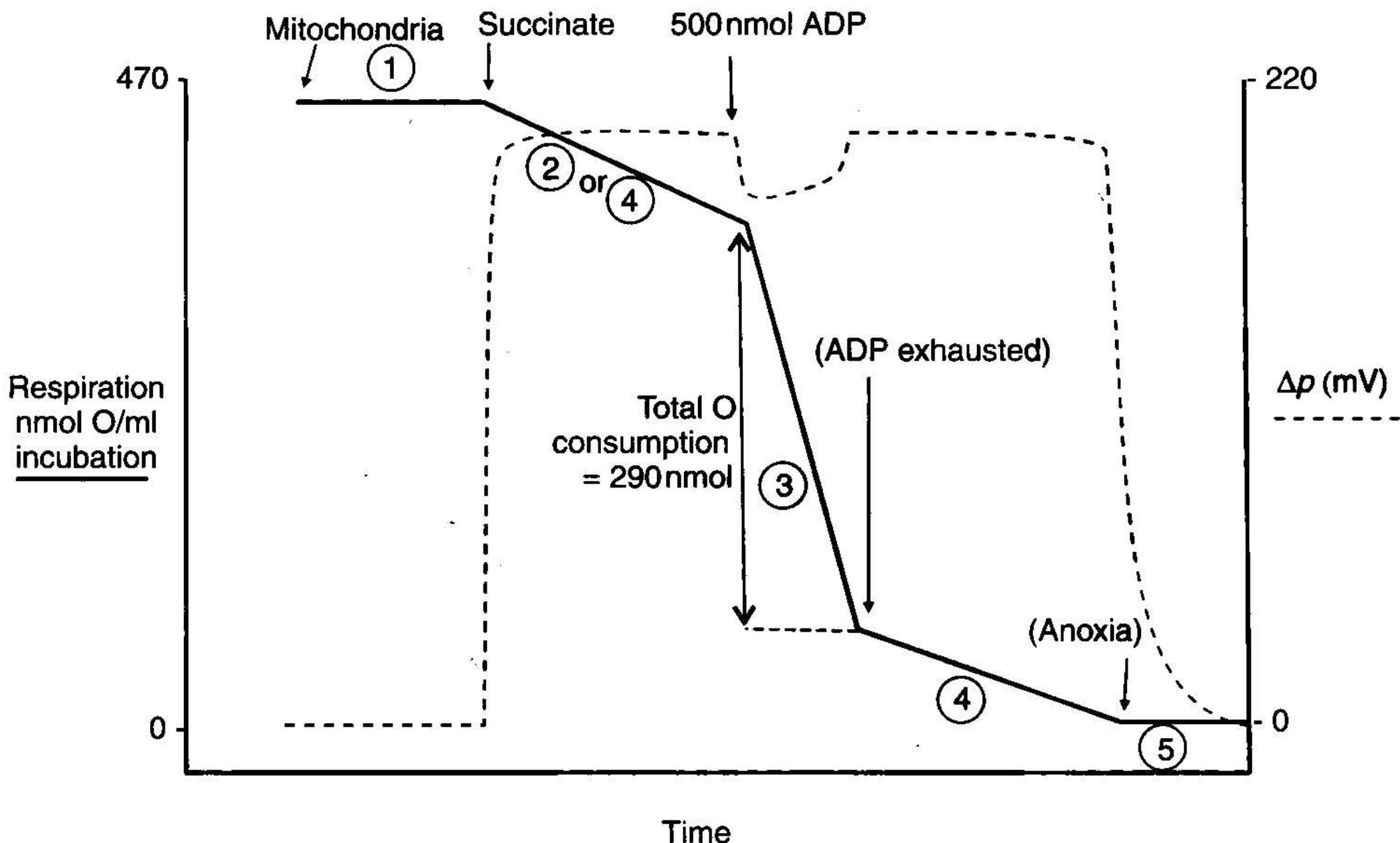
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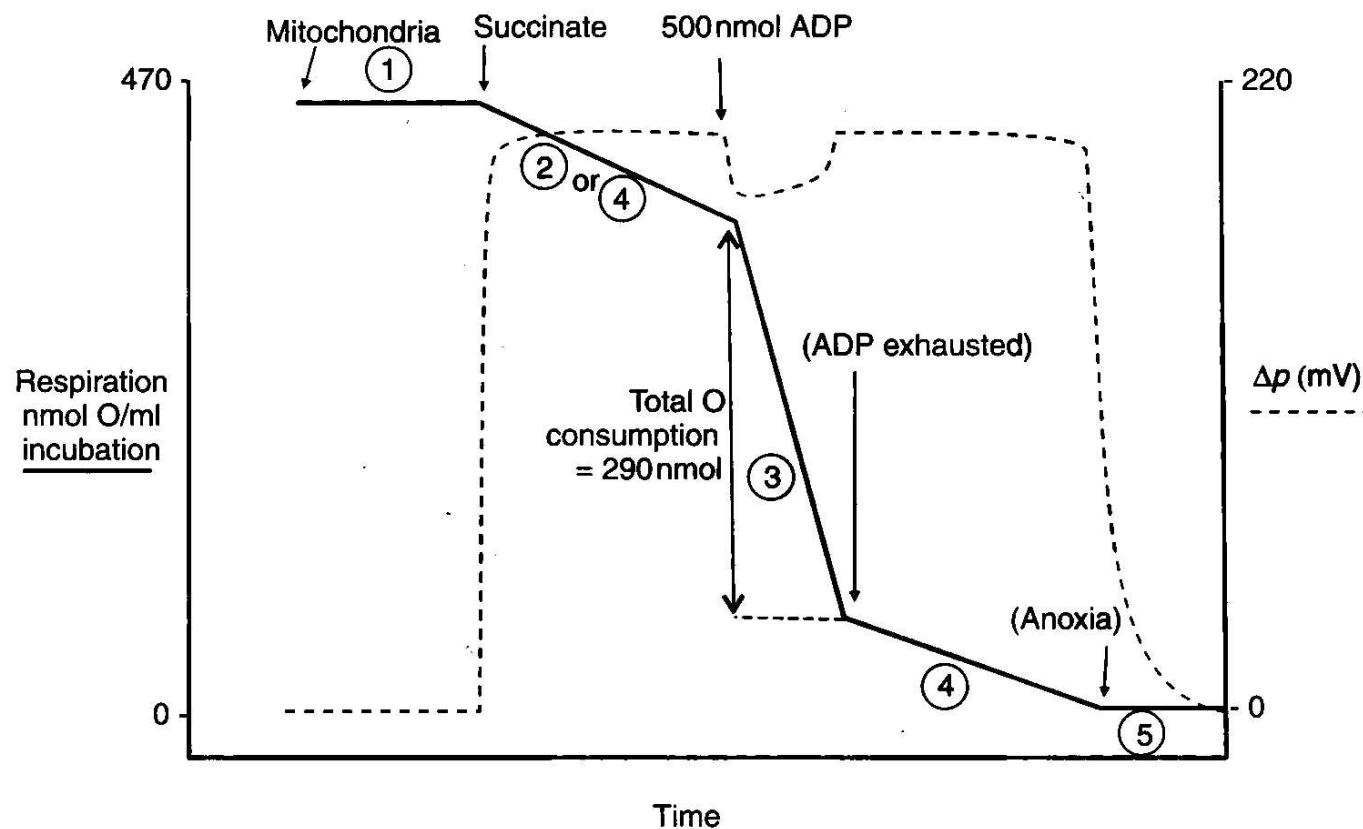
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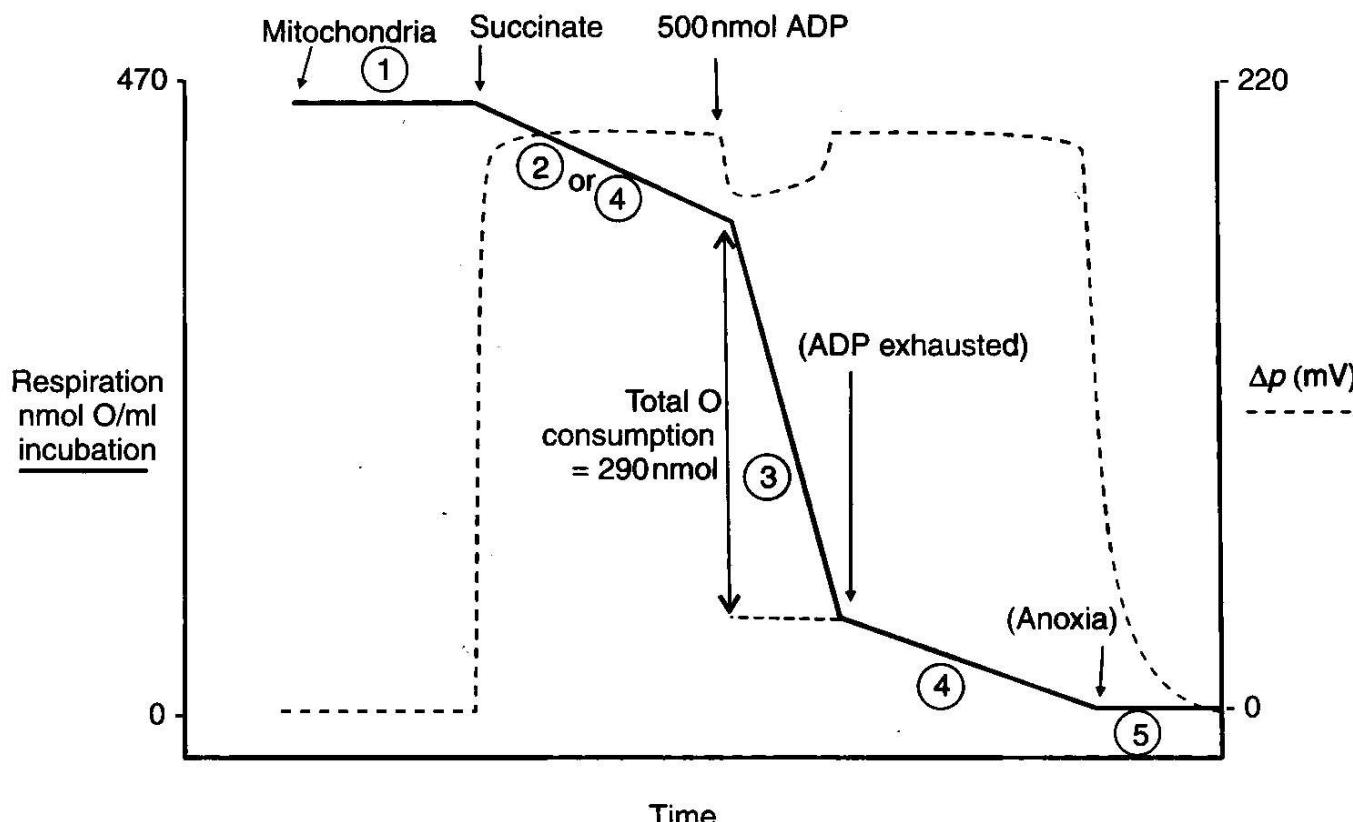
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- c) The rate then remains high until all the ADP has been phosphorylated to give ATP. At this point the rate declines again to state IV







**Figure 4.9 Respiratory 'states' and the determination of P/O ratios.**

In this experiment mitochondria were added to an oxygen electrode chamber, followed by succinate as substrate. Respiration is slow as the proton circuit is not completed by  $H^+$  re-entry through the ATP synthase. That there is any respiration at all is because of a slow proton leak across the membrane. A limited amount of ADP is added, allowing the ATP synthase to synthesize ATP coupled to proton re-entry across the membrane, 'state 3' (Section 4.5). When this is exhausted, respiration slows and finally anoxia is attained. The circled numbers refer to the respiratory 'states'. If the amount of ADP is known, the oxygen uptake during the accelerated 'state 3' respiration can be quantified allowing a P/O ratio to be calculated (moles ATP synthesized per mol O). Since the proton leak is almost negligible in 'state 3' (Section 4.6.2), the total oxygen uptake during 'state 3' is effectively used for ATP synthesis. In this example, the ADP/O ratio for the substrate is found to be  $500/290 = 1.72$ . Note the bioenergetic convention of referring to 'O', i.e.  $\frac{1}{2}O_2$ , which is equivalent to  $2e^-$ . Also, the controlled respiration prior to addition of ADP, which is strictly termed 'state 2' is functionally the same as state 4, and the latter term is usually used for both states. The dotted trace reports the values of  $\Delta p$  during the experiment.

# Characteristics of oxidative phosphorylation

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- 2] Respiratory control
- 3] Uncoupling agents
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Uncouplers dissipate this energised state as heat, and so no ATP synthesis takes place. Electron transfer then proceeds at a maximal rate because it is no longer controlled by back-pressure from the energised state – respiratory control has been abolished.

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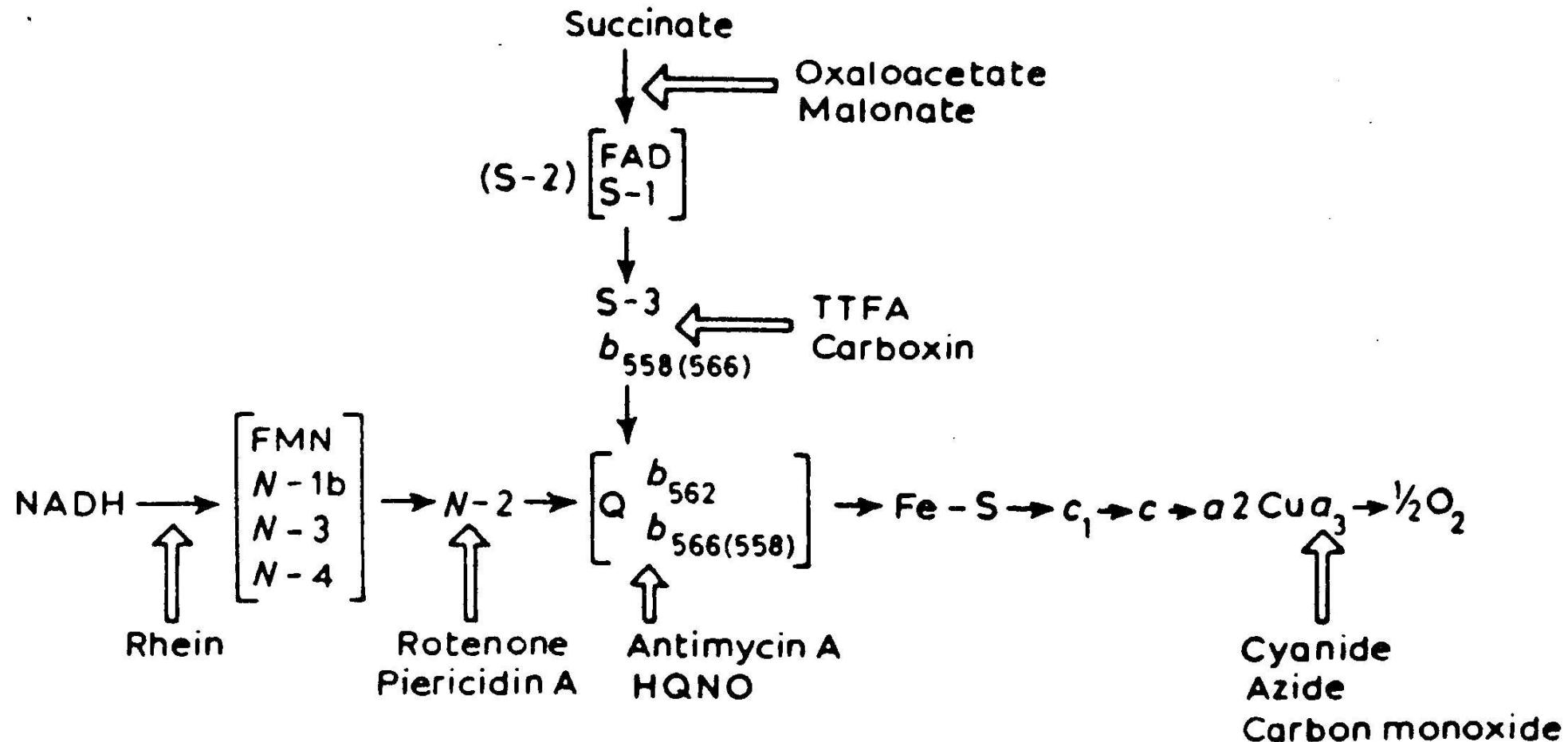
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This evidence indicates that uncouplers act between electron transfer and ATP synthesis, that is, on the energised state.

A view of mitochondrial electron transfer as a series of redox (oxidation-reduction) carriers in the inner mitochondrial membrane. This view concentrates on redox centres, substrates, and inhibitors, and neglects the significance of the proteins to which they bind, and of the mitochondrial inner membrane to which the proteins are bound.



**Fig. 2.19** The mitochondrial respiratory chain.



Membrane Biochemistry

Next lecture....

# ATP Synthase – Coupling ATPase

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